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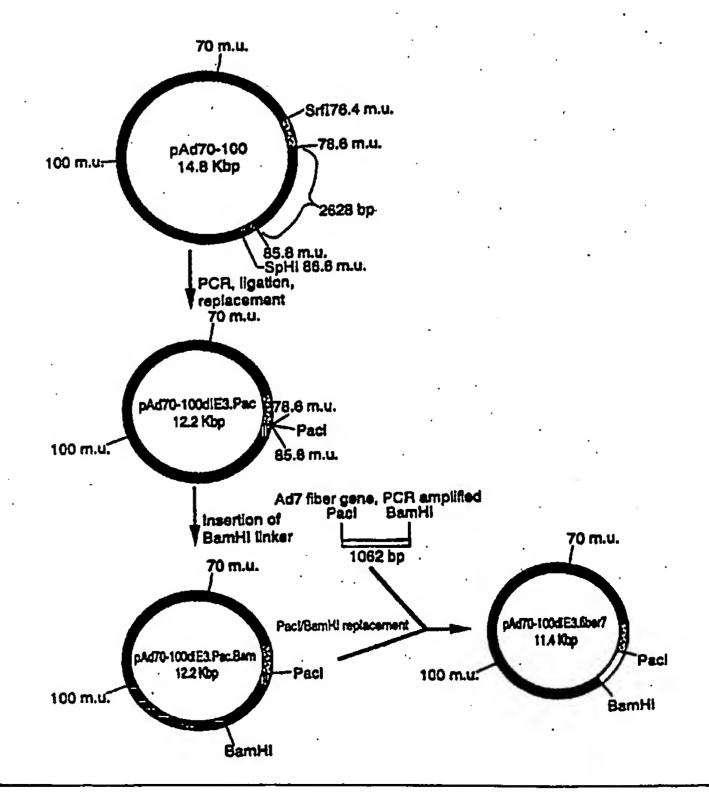
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(54) Title: CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME

(57) Abstract

The present invention provides a chimeric adenoviral coat protein (particularly a chimeric adenovirus hexon protein). The chimeric adenovirus coat protein has a decreased ability or inability to be recognized by a neutralizing antibody directed against the corresponding wild-type adenovirus coat protein.



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CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME

TECHNICAL FIELD OF THE INVENTION

The present invention relates to a chimeric adenoviral coat protein and a recombinant adenovirus comprising same. In particular, the invention provides a chimeric adenoviral hexon protein and a recombinant adenovirus comprising the chimeric adenoviral hexon protein. Such a recombinant adenovirus can be employed inter alia in gene therapy.

BACKGROUND OF THE INVENTION

In vivo gene therapy is a strategy in which nucleic acid, usually in the form of DNA, is administered to modify the genetic repertoire of target cells for therapeutic purposes. This can be accomplished efficiently using a recombinant adenoviral vector encoding a so-called "therapeutic gene". A therapeutic gene is generally considered a gene that corrects or compensates for an underlying protein deficit or, alternately, a gene that is capable of down-regulating a particular gene, or counteracting the negative effects of its encoded product, in a given disease state or syndrome. Recombinant adenoviral vectors have been used to transfer one or more recombinant genes to diseased cells or tissues in need of treatment. As reviewed by Crystal, Science, 270, 404-410 (1995), such vectors are preferred over other vectors commonly employed for gene therapy (e.g., retroviral vectors) since adenoviral vectors can be produced in high titers (i.e., up to 1013 viral particles/ml), and they efficiently transfer genes to nonreplicating, as well as replicating, cells. Moreover, adenoviral vectors are additionally preferred based on their normal tropism for

the respiratory epithelium in cases where the targeted tissue for somatic gene therapy is the lung, as well as for other reasons (see, e.g., Straus, In Adenoviruses, Plenan Press, New York, NY, 451-496 (1984)); Horwitz et al., In Virology, 2nd Ed., Fields et al., eds., Raven Press, New York, NY, 1679-1721 (1990); Berkner, BioTechniques, 6, 616 (1988); Chanock et al., JAMA, 195, 151 (1966); Haj-Ahmad et al., J. Virol., 57, 267 (1986); and Ballay et al., EMBO, 4, 3861 (1985)).

There are 49 human adenoviral serotypes, categorized into 6 subgenera (A through F) based on nucleic acid comparisons, fiber protein characteristics, and biological properties (Crawford-Miksza et al., J. Virol., 70, 1836-1844 (1996)). The group C viruses (e.g., serotypes 2 and 5, or Ad2 and Ad5) are well characterized. It is these serotypes that currently are employed for gene transfer studies, including human gene therapy trials (see, e.g., Rosenfeld et al., Science, 252, 431-434 (1991); Rosenfeld et al., Cell, 68, 143-155 (1992); Zabner, Cell, 75, 207-216 (1993); Crystal et al., Nat. Gen., 8, 42-51 (1994); Yei et al., Gene Therapy, 1, 192-200 (1994); Chen et al., Proc. Natl. Acad. Sci., 91, 3054-3057 (1994); Yang et al., Nat. Gen., 7, 362-369 (1994); Zabner et al., Nat. Gen., 6, 75-83 (1994)). Other groups and serotypes include, but are not limited to: group A (e.g., serotypes 12 and 31), group B (e.g., serotypes 3 and 7), group D (e.g., serotypes 8 and 30), group E (e.g., serotype 4) and group F (e.g., serotypes 40 and 41) (Horwitz et al., supra).

In terms of general structure, all adenoviruses examined to date are nonenveloped, regular icosahedrons of about 65 to 80 nanometers in diameter. Adenoviruses are comprised of linear, double-stranded DNA that is complexed with core proteins and surrounded by the adenoviral capsid. The capsid is comprised of 252 capsomeres, of which 240 are hexons and 12 are pentons. The hexon

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capsomere provides structure and form to the capsid (Pettersson, in <u>The Adenoviruses</u>, pp. 205-270, Ginsberg, ed., (Plenum Press, New York, NY, 1984)), and is a homotrimer of the hexon protein (Roberts et al., <u>Science</u>, <u>232</u>, 1148-1151 (1986)). The penton comprises a penton base, which is bound to other hexon capsomeres, and a fiber, which is noncovalently bound to, and projects from, the penton base. The penton fiber protein comprises three identical polypeptides (i.e., polypeptide IV). The Ad2 penton base protein comprises five identical polypeptides (i.e., polypeptide III) of 571 amino acids each (Boudin et al., Virology, 92, 125-138 (1979)).

The adenoviruses provide an elegant and efficient means of transferring therapeutic genes into cells. However, one problem encountered with the use of adenoviral vectors for gene transfer in vivo is the generation of antibodies to antigenic epitopes on adenoviral capsid proteins. If sufficient in titer, the antibodies can limit the ability of the vector to be used more than once as an effective gene transfer vehicle. For instance, animal studies demonstrate that intravenous or local administration (e.g., to the lung, heart or peritoneum) of an adenoviral type 2 or 5 gene transfer vector can result in the production of antibodies directed against the vector which prevent expression from the same serotype vector administered 1 to 2 weeks later (see, e.g., Yei et al., supra; Zabner (1994), supra; Setoguchi et al., Am. J. Respir. Cell. Mol. Biol., 10, 369-377 (1994); Kass-Eisler et al., Gene Therapy, 1, 395-402 (1994); Kass-Eisler et al., Gene Therapy 3, 154-162 (1996)). This is a drawback in adenoviral-mediated gene therapy, since many uses of an adenoviral vector (e.g., for prolonged gene therapy) require repeat administration inasmuch as the vector does not stably integrate into the host cell genome. The mechanism by which antibodies

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directed against an adenovirus are able to prevent or reduce expression of an adenoviral-encoded gene is unclear. However, the phenomenon is loosely referred to as "neutralization", and the responsible antibodies are termed "neutralizing antibodies."

There are three capsid structures against which neutralizing antibodies potentially can be elicited: fiber, penton, and hexon (Pettersson, supra). The hexon protein, and to a lesser extent the fiber protein, comprise the main antigenic determinants of the virus, and also determine the serotype specificity of the virus (Watson et al., J. Gen. Virol., 69, 525-535 (1988); Wolfort et al., J. Virol., 62, 2321-2328 (1988); Wolfort et al., J. Virol., 56, 896-903 (1985); Crawford-Miksza et al., supra). Researchers have examined and compared the structure of these coat proteins of different adenoviral serotypes in an effort to define the regions of the proteins against which neutralizing antibodies are elicited.

The Ad2 hexon trimer is comprised of a pseudohexagonal base and a triangular top formed of three towers (Roberts et al., supra; Athappilly et al., J. Mol. Biol., 242, 430-455 (1994)). The base pedestal consists of two tightly packed eight-stranded antiparallel beta barrels stabilized by an internal loop. The predominant regions in hexon protein against which neutralizing antibodies are directed appear to be in loops 1 and 2 (i.e., LI or 11, and LII or 12, respectively) in one of the three towers. For instance, Kinloch et al. (J. Biol. Chem., 258, 6431-6436 (1984)) compared adenoviral hexon sequences and theorized that the serotype-specific antigenic determinants on hexon are located in amino acid residues 120 to 470 encompassing the 11 and 12 loops since type-specific sequence differences are mainly concentrated in this region. Toogood et al. (J. Gen.

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<u>Virol.</u>, <u>73</u>, 1429-1435 (1992)) used peptides from this region to generate specific anti-loop antisera and confirmed that antibodies against residues 281-292 of 11 and against residues 441-455 of 12 were sufficient to neutralize infection. Also, Crompton et al. (<u>J. Gen. Virol.</u>, <u>75</u>, 133-139 (1994)) modified these loops to accept neutralizing epitopes from polio virus, and demonstrated that infection with the resultant adenoviral vector generated neutralizing immunity against polio virus. More recently it was demonstrated that the hexon protein is composed of seven discrete hypervariable regions in loops and 1 and 2 (HVR1 to HVR7) which vary in length and sequence between adenoviral serotypes (Crawford-Miksza et al., <u>supra</u>).

Less is known regarding the regions of the fiber protein against which neutralizing antibodies potentially can be directed. However, much data is available on the structure of the fiber protein. The trimeric fiber protein consists of a tail, a shaft, and a knob (Devaux et al., J. Molec. Biol., 215, 567-588 (1990)). The fiber shaft region is comprised of repeating 15 amino acid motifs, which are believed to form two alternating beta strands and beta bends (Green et al., EMBO J., 2, 1357-1365 (1983)). The overall length of the fiber shaft region and the number of 15 amino acid repeats differ between adenoviral serotypes. The receptor binding domain of the fiber protein and sequences necessary for fiber trimerization are localized in the knob region encoded by roughly the last 200 amino acids of the protein (Henry et al., J. Virol., 68(8), 5239-5246 (1994)); Xia et al., Structure, 2(12), 1259-1270 (1994)). Furthermore, all adenovirus serotypes appear to possess a type of specific moiety located in the knob region (Toogood et al., supra.)

Given the existence of these potential epitopes in hexon protein and fiber protein, it is understandable

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that, in some cases, difficulties have been encountered using adenovirus as a vector for gene therapy.

Accordingly, recombinant adenoviral vectors capable of escaping such neutralizing antibodies (in the event they are preexisting and hamper gene expression commanded by adenovirus in an initial dose), and which would allow repeat doses of adenoviral vectors to be administered, would significantly advance current gene therapy methodology.

Thus, the present invention seeks to overcome at least some of the aforesaid problems of recombinant adenoviral gene therapy. In particular, it is an object of the present invention to provide a recombinant adenovirus comprising a chimeric coat protein that has a decreased ability or inability to be recognized by antibodies (i.e., neutralizing antibodies) directed against the corresponding wild-type adenovirus coat protein. These and other objects and advantages of the present invention, as well as additional inventive features, will be apparent from the description of the invention provided herein.

BRIEF SUMMARY OF THE INVENTION

The present invention provides a chimeric adenovirus coat protein (particularly a chimeric adenovirus hexon protein) comprising a nonnative amino acid sequence. The chimeric adenovirus coat protein is not recognized by, or has a decreased ability to be recognized by, a neutralizing antibody directed against the corresponding wild-type (i.e., native) coat protein. The chimeric adenovirus coat protein enables a vector (such as an adenovirus) comprising the corresponding protein to be administered repetitively, or to be administered following administration of an adenovirus vector comprising the corresponding wild-type coat protein. It also enables a

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vector (such as an adenovirus) comprising the chimeric protein to be administered and effect gene expression in the case where there are preexisting neutralizing antibodies directed against the wild-type adenovirus coat protein. The present invention also provides a vector, particularly an adenoviral vector, that comprises a chimeric adenovirus coat protein such as chimeric adenovirus hexon protein (and which optionally further comprises a chimeric adenovirus fiber and/or penton base protein), and methods of constructing and using such a vector.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a diagram of the method employed to construct the vector pAd70-100dlE3.fiber7.

Figure 2 is a partial restriction map of the vector pGBS.59-100(HSF:RGD).

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides, among other things, a chimeric adenovirus coat protein. The chimeric adenovirus coat protein comprises a nonnative amino acid sequence, such that the chimeric adenovirus coat protein (or a vector comprising the chimeric adenovirus coat protein) has a decreased ability or inability to be recognized by antibodies (e.g., neutralizing antibodies) directed against the corresponding wild-type adenovirus coat protein.

Chimeric Adenovirus Coat Protein

A "coat protein" according to the invention is either an adenoviral penton base protein, an adenoviral hexon protein, or an adenoviral fiber protein. Preferably a coat protein is a adenoviral hexon protein or an adenoviral fiber protein. Any one of the serotypes of

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human or nonhuman adenovirus can be used as the source of the coat protein, or its gene or coding sequence.

Optimally, however, the adenovirus coat protein is that of a Group B or C adenovirus and, preferably, is that of Adl, Ad2, Ad3, Ad5, Ad6, Ad7, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, or Ad48.

The chimeric adenovirus coat protein (or a vector, such as adenoviral vector, comprising the chimeric adenovirus coat protein) has a decreased ability or an inability to be recognized by an antibody (e.g., a neutralizing antibody) directed against the corresponding wild-type adenovirus coat protein. A "neutralizing antibody" is an antibody that either is purified from or is present in serum. As used herein, an antibody can be a single antibody or a plurality of antibodies. An antibody is "neutralizing" if it inhibits infectivity of (i.e., cell entry) or gene expression commanded by an adenovirus comprising wild-type coat protein, or if it exerts a substantial deleterious effect on infectivity of or gene expression commanded by an adenovirus comprising wild-type coat protein, as compared, for instance, to any effect on any other adenoviral property.

An ability or inability of a chimeric coat protein to "be recognized by" (i.e., interact with) a neutralizing antibody directed against the wild-type adenovirus coat protein can be assessed by a variety of means known to those skilled in the art. For instance, the removal of one or more epitopes for a neutralizing antibody present in a wild-type adenovirus coat protein to generate a chimeric adenovirus coat protein will result in a decreased ability or inability of the chimeric coat protein to be recognized by the neutralizing antibody. Also, such a decreased ability or inability to interact with a neutralizing antibody directed against wild-type coat protein can be demonstrated by means of a

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neutralization test (see, e.g., Toogood et al., <u>supra;</u> Crawford-Miksza et al., <u>supra;</u> Mastrangeli et al., <u>Human</u> <u>Gene Therapy</u>, <u>7</u>, 79-87 (1996)), or as further described herein.

Generally, an "inability" of a chimeric adenovirus coat protein (or a vector comprising a chimeric adenovirus coat protein) to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein means that such an antibody does not interact with the chimeric coat protein, and/or exhibits no substantial deleterious effect on infectivity of or gene expression commanded by an adenovirus comprising wild-type coat protein, as compared, for instance, to any effect on any other adenoviral property.

A "decreased ability" to be recognized by neutralizing antibody directed against wild-type adenovirus coat protein refers to any decrease in the ability of the chimeric adenovirus coat protein (or a vector comprising the chimeric coat protein) to be recognized by an antibody directed against the corresponding wild-type adenovirus coat protein as compared to the wild-type adenovirus coat protein. When such ability/inability is assessed by means of a neutralization test in particular, preferably a "decreased ability" to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein is exhibited by from about a 10% to about a 99% increase in the ability of a recombinant adenovirus comprising the chimeric coat protein to cause a visible cytopathic effect (c.p.e.) in cells such as A549 cells or COS-1 cells (or other such cells appropriate for a neutralization assay) in the presence of the neutralizing antibody as compared to an adenovirus comprising the wild-type coat protein against which the neutralizing antibody is directed.

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Furthermore, a decreased ability or inability of an adenovirus chimeric coat protein (or a vector comprising a chimeric adenovirus coat protein) to interact with a neutralizing antibody can be shown by a reduction of inhibition (from about 10% to about 99%) or no inhibition at all of cell infectivity by a recombinant vector (such as an adenoviral vector) containing the chimeric coat protein as compared to a recombinant vector containing the wild-type protein. Also, a decreased ability or inability of an adenovirus chimeric coat protein (or a vector comprising a chimeric adenovirus coat protein) to interact with a neutralizing antibody can be shown by a reduction of inhibition (from about 10% to about 99%) or no inhibition at all of gene expression commanded by a recombinant vector (such as an adenoviral vector) containing the chimeric coat protein as compared to a recombinant vector containing the wild-type coat protein. These tests can be carried out when the recombinant adenovirus containing the chimeric coat protein is administered following the administration of an adenovirus containing the wild-type coat protein, or when the recombinant adenovirus is administered to a host that has never before encountered or internalized adenovirus (i.e., a "naïve" host). These methods are described, for instance, in the Examples which follow as well as in . Mastrangeli et al., supra. Other means such as are known to those skilled in the art also can be employed.

The coat protein is "chimeric" in that it comprises a sequence of amino acid residues that is not typically found in the protein as isolated from, or identified in, wild-type adenovirus, which comprises the so-called native coat protein, or "wild-type coat protein". The chimeric coat protein thus comprises (or has) a "nonnative amino acid sequence". By "nonnative amino acid sequence" is meant any amino acid sequence (i.e., either component

residues or order thereof) that is not found in the native coat protein of a given serotype of adenovirus, and which preferably is introduced into the coat protein at the level of gene expression (i.e., by production of a nucleic acid sequence that encodes the nonnative amino acid sequence). Generally, the nonnative amino acid sequence can be obtained by deleting a portion of the amino acid sequence, deleting a portion of the amino acid sequence and replacing the deleted amino sequence with a so-called "spacer region", or introducing the spacer region into an unmodified coat protein. Preferably such manipulations result in a chimeric adenovirus coat protein according to the invention that is capable of carrying out the functions of the corresponding wild-type adenovirus coat protein (or, at least that when incorporated into an adenovirus, will allow appropriate virion formation and will not preclude adenoviral-mediated cell entry), and, optimally, that is not impeded in its proper folding. Also, it is desirable that the manipulations do not result in the creation of new epitopes for differing antibodies, unless, of course, such epitopes do not interfere with use of an adenovirus containing the chimeric coat protein as a gene transfer vehicle in vivo.

In particular, a nonnative amino acid sequence according to the invention preferably comprises a deletion of a region of a wild-type adenovirus coat protein, particularly an adenovirus hexon or fiber protein.

Optimally the resultant nonnative amino acid sequence is such that one or more of the existing epitopes for neutralizing antibodies directed against the corresponding wild-type adenovirus coat protein have been rendered non-immunogenic. Desirably, the region deleted comprises from about 1 to about 750 amino acids, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the

region deleted comprises a smaller region less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. The chimeric coat protein also desirably comprises a plurality of such deletions. Thus, according to the invention, the chimeric adenovirus coat protein comprises modification of one or more amino acids, and such modification is made in one or more regions.

In a preferred embodiment of the present invention, a nonnative amino acid sequence comprises a deletion of one or more regions of a wild-type adenovirus hexon protein, wherein preferably the hexon protein is the Ad2 hexon protein [SEQ ID NO:2] (which is encoded by the sequence of SEQ ID NO:1; GenBank® Data Bank Accession Number U20821), or the Ad5 hexon protein [SEQ ID NO:3] (GenBank® Data Bank Accession Number M73260, which is encoded by the sequence of SEQ ID NO:4), or the Ad7 hexon protein (GenBank® Data Bank Accession Number x76551). Alternately, preferably the hexon protein is the protein sequence reported by Crawford-Miksza et al. (Ad2 hexon [SEQ ID NO:52], Ad5 hexon SEQ ID NO:54]). In particular, the sequences of Crawford-Miksza et al. differ over those reported in the GenBank® Data Bank in that the amino acid residue reported as the first in the Crawford-Miksza et al. sequences is not Met, and the Ad5 hexon sequence is reported as terminating with "Gln His" instead of with "Thr Thr". As employed herein, the numbering of adenovirus hexon amino acid residues corresponds to that in Crawford-Miksza et al.

Desirably the region(s) of the deletion comprises an internal hexon protein sequence ("internal" meaning not at or near the C- or N-terminus of the protein; "near" referring to a distance of 500 amino acids or less), preferably a hypervariable region, e.g., as reported in Crawford-Miksza et al. In particular, optimally, the

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internal region of the wild-type hexon protein that is deleted to generate the chimeric hexon protein comprises the entirety of 11 loop, preferably from about residue 131 to about residue 331 of the Ad2 hexon protein [SEQ ID NO:6] (which is encoded by the sequence of SEQ ID NO:5), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad5 [SEQ ID NO:8] (which is encoded by the sequence of SEQ ID NO:7), Ad6, Ad7, Ad8, Ad12, Ad16, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

Alternately, preferably the internal region of the wild-type hexon protein that is deleted to produce the chimeric hexon protein comprises one or more regions (e.g., smaller regions) of the 11 loop. Optimally the region deleted comprises a hypervariable region. Desirably the one or more regions of the 11 loop deleted are regions (i.e., hypervariable regions) selected from this group consisting of the HVR1 region, the HVR2 region, the HVR3 region, the HVR4 region, the HVR5 region, and the HVR6 region. Moreover, preferably the region of the wildtype protein that is deleted (or otherwise manipulated as described herein) occurs on the external surface of the hexon protein. Thus, HVR2, HVR3, HVR4, and HVR5 -- each of which are externally located regions of the hexon protein -- are particularly preferred for deletion or modification.

The "HVR1 region" preferably comprises from about amino acid 137 to about amino acid 188 of the Ad2 hexon protein [SEQ ID NO:10] (which is encoded by the sequence of SEQ ID NO:9), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:12] (which is encoded by the sequence of SEQ ID NO:11), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48,

BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR2 region" preferably comprises from about amino acid 194 to about amino acid 204 of the Ad2 hexon protein [SEQ ID NO:14] (which is encoded by the sequence of SEQ ID NO:13), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:16] (which is encoded by the sequence of SEQ ID NO:15), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR3 region" preferably comprises from about amino acid 222 to about amino acid 229 of the Ad2 hexon protein [SEQ ID NO:18] (which is encoded by the sequence of SEQ ID NO:17), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:20] (which is encoded by the sequence of SEQ ID NO:19), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR4 region" preferably comprises from about amino acid 258 to about amino acid 271 of the Ad2 hexon protein [SEQ ID NO:22] (which is encoded by the sequence of SEQ ID NO:21), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:24] (which is encoded by the sequence of SEQ ID NO:23), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR5 region" preferably comprises from about amino acid 278 to about amino acid 294 of the Ad2 hexon protein [SEQ ID NO:26] (which is encoded by the sequence

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of SEQ ID NO:25), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:28] (which is encoded by the sequence of SEQ ID NO:27), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. In particular, preferably the deleted region comprises from about amino acid 297 to about amino acid 304 just outside of the HVR5 region of the Ad2 hexon protein [SEQ ID NO:30] (which is encoded by the sequence of SEQ ID NO:29), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:32] (which is encoded by the sequence of SEQ ID NO:31), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR6 region" preferably comprises from about amino acid 316 to about amino acid 327 of the Ad2 hexon protein [SEQ ID NO:34] (which is encoded by the sequence of SEQ ID NO:33), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:36] (which is encoded by the sequence of SEQ ID NO:35), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

In another preferred embodiment of the invention, the internal region of the wild-type hexon protein that is deleted to generate the chimeric hexon protein comprises the entirety of the 12 loop, preferably from about residue 423 to about residue 477 of the Ad2 hexon protein [SEQ ID NO:38] (which is encoded by the sequence of SEQ ID NO:37), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from

Ad1, Ad3, Ad5 [SEQ ID NO:40] (which is encoded by the sequence of SEQ ID NO:39), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. Alternately, preferably the internal region of the wild-type hexon protein that is deleted to produce the chimeric hexon protein comprises one or more smaller regions (e.g., hypervariable regions) of the 12 loop. In particular, preferably the smaller region of the 12 loop comprises the HVR7 region.

The "HVR7 region" preferably comprises from about amino acid 433 to about amino acid 465 of the Ad2 hexon protein [SEQ ID NO:42] (which is encoded by the sequence of SEQ ID NO:41), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:44] (which is encoded by the sequence of SEQ ID NO:43), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza In particular, preferably the deleted et al., supra. region comprises from about amino acid 460 to about amino acid 466 of the HVR7 region (i.e., extending one base pair outside of this region) of the Ad2 hexon protein [SEQ ID NO:46] (which is encoded by the sequence of SEQ ID NO:45), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:48] (which is encoded by the sequence of SEQ ID NO:47), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

Along the same lines, the chimeric adenovirus hexon protein desirably comprises deletions in one or both of the aforementioned regions, i.e., the hexon protein comprises deletions in one or both of the 11 and 12 loops,

which deletions can constitute the entirety of the loop(s), or can comprise deletions of one or more smaller regions (e.g., hypervariable regions) in one or both of the hexon loops. In particular, desirably the deleted region(s) are selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

An "equivalent" is a naturally occurring variation of an amino acid or nucleic acid sequence, e.g., as are observed among different strains of adenovirus. A conservative variation is a variation of an amino acid sequence that results in one or more conservative amino acid substitution(s). A "conservative amino acid substitution" is an amino acid substituted by an alternative amino acid of similar charge density, hydrophilicity/hydrophobicity, size, and/or configuration (e.g., basic, Arg and Lys; aliphatic Ala, Cys, Gly, Ile, Leu, Met and Val; aromatic, Phe, Tyr, Trp, and His; hydrophilic, Glu, Gln, Asn, and Asp; hydroxyl, Ser and Thr).

In another preferred embodiment, the nonnative amino acid sequence of the chimeric adenoviral coat protein (i.e., particularly a chimeric adenoviral fiber or hexon protein) comprises a deletion of one or more region(s) of the wild-type adenovirus coat protein (particularly the 11)

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and/or 12 loops, and, most particularly, the HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and/or HVR7 regions of the wildtype adenovirus hexon protein) as previously described, and further comprises a replacement of the region(s) with a spacer region preferably of from 1 to about 750 amino acids, especially of from about 1 to about 500 amino acids, and particularly of from about 1 to about 300 amino acids. It also is desirable that the region deleted and replaced comprises a smaller region less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. chimeric coat protein also desirably comprises a plurality of such replacements. Thus, according to the invention, the chimeric adenovirus coat protein comprises modification of one or more amino acids, and such modification is made in one or more regions which can be a smaller region. A spacer region of the aforementioned size also preferably simply can be inserted into one of the aforementioned regions (particularly into the 11 and/or 12 loop, or one or more of the aforementioned HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and HVR7 regions of the adenovirus hexon protein) in the absence of any deletion to render the resultant chimeric protein nonimmunogenic by, for instance, destroying the ability of a neutralizing antibody to interact with that particular site (e.g., by changing the spatial juxtaposition of critical amino acids with which the antibody interacts).

Optimally the spacer region comprises a nonconservative variation of the amino acid sequence of wild-type adenovirus coat protein (particularly wild-type adenovirus hexon protein) that comprises an epitope for a neutralizing antibody, and which may or may not be deleted upon the insertion of the spacer region. A "nonconservative variation" is a variation of this amino acid sequence that does not result in the creation or

recreation in the chimeric adenovirus coat protein of the epitope for a neutralizing antibody directed against the wild-type adenovirus coat protein, and, in particular, is a variation of the spacer region that results in one or more nonconservative amino acid insertion(s) or substitution(s) in this region. A "nonconservative amino acid substituted by an alternative amino acid of differing charge density, hydrophilicity/hydrophobicity, size, and/or configuration (e.g., a change of a basic amino acid for an acidic amino acid, a hydrophilic amino acid for a hydrophobic amino acid, and the like).

Desirably the spacer region does not interfere with the functionality of the chimeric adenovirus coat protein, particularly the chimeric adenovirus hexon or fiber protein, e.g., the ability of hexon protein to bind penton base protein or other hexon capsomeres, or the ability of penton fiber to bind penton base and/or to a cell surface receptor. Such functionality can be assessed by virus viability. Similarly, the absence of the creation or recreation of the epitope(s) for a neutralizing antibody directed against the wild-type coat (e.g., hexon and/or fiber) protein can be confirmed using techniques as described in the Examples which follow (e.g., by ensuring the antibody, which may be in a carrier fluid such as serum or other liquid, binds the wild-type adenovirus coat protein, but not the chimeric adenovirus coat protein).

Preferably the spacer region incorporated into the adenovirus coat protein (i.e., either as an insertion into the wild-type coat protein, or to replace one or more deleted region(s) of the wild-type adenovirus coat protein) comprise a series of polar and/or charged amino acids (e.g., Lys, Arg, His, Glu, Asp, and the like), or amino acids with intermediate polarity (e.g., Gln, Asn, Thr, Ser, Met, and the like). In particular, desirably

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the spacer region comprises the sequence of SEQ ID NO:50 (which is encoded by the sequence of SEQ ID NO:49), and equivalents and conservative variations of SEQ ID NO:50. Alternately, the spacer region can comprise any other sequence like the FLAG octapeptide sequence of SEQ ID NO:50 that will not interfere with the functionality of the resultant chimeric protein.

In still yet another preferred embodiment, a region of a wild-type adenovirus coat protein (particularly an adenovirus hexon and/or fiber protein) is deleted and replaced with a spacer region comprising the corresponding. coat protein region of another adenoviral serotype. Preferably in this embodiment the spacer region is of a different adenoviral group. For instance, preferably a region of an Ad2 coat protein can be replaced with the corresponding region of an Ad5 or Ad7 coat protein (or any other serotype of adenovirus as described above), and vice versa. It also is preferable that such a spacer region comprising the coat protein region of another adenoviral serotype is simply inserted into the corresponding coat protein region of the chimeric coat protein. case, the likelihood of obtaining a chimeric hexon protein that is functional can be increased by making sure that the size of the hypervariable domain resulting from such insertion approximates the size of a known hypervariable domain. For instance, the HVR1 region of Ad40 is about 30 amino acids smaller than the HVR1 region of Ad2 (as well as other adenoviruses such as Ad5, Ad8, etc.). preferably a spacer region of about 30 amino acids can be incorporated into the Ad40 HVR1 region to produce a chimeric adenovirus hexon protein. In particular, desirably the region of Ad2 (or other adenovirus) that is not present in Ad40 (i.e., approximately amino acid residues 138 to 174), or a portion thereof, is introduced

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into Ad40 to produce the chimeric adenoviral hexon protein.

According to the invention, desirably the nonnative amino acid sequence of a chimeric coat protein comprises a plurality of such replacements or insertions. When the coat protein is incorporated into an adenoviral vector, preferably the entire coat protein of one adenoviral serotype can be substituted with the entire coat protein of another adenoviral serotype, as described further herein.

The region or regions of wild-type adenovirus hexon protein that are deleted and replaced by the spacer region, or into which the spacer region is inserted, can be any suitable region(s) and desirably comprise one or more of the regions described above with respect to the hexon protein deletions. For instance, preferably the one or more regions into which the spacer region is inserted or which the spacer region replaces comprises the entirety of the 11 and/or 12 loop, or a sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

Similarly, the spacer region itself (i.e., both for insertion as well as replacement) preferably comprises the entirety of the 11 and/or 12 loop, or a sequence selected

from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:34, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

The fiber protein also preferably is altered in a similar fashion as described for modification of hexon protein to escape antibodies directed in particular against wild-type adenovirus fiber protein. Fiber protein sequences and methods of modifying fiber protein are known to those skilled in the art (see, e.g., Xia et al., supra; Novelli et al., Virology, 185, 365-376 (1991)). The fiber manipulations can be carried out in the absence of, or along with, modifications to the adenovirus hexon protein. In particular, preferably the fiber protein can be replaced in its entirety, or in part, with sequences of a fiber protein from a different serotype of adenovirus. Also, preferably, deletions can be made of fiber sites that constitute an epitope for a neutralizing antibody, and/or insertions can be made at the site to destroy the ability of the protein to interact with the antibody.

Nucleic Acid Encoding The Chimeric Adenovirus Coat Protein

Preferably the chimeric adenovirus coat protein (particularly the chimeric adenovirus hexon or fiber protein) comprises a nonnative amino acid sequence wherein

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the alteration is made at the level of DNA. invention preferably provides an isolated and purified nucleic acid encoding a chimeric adenovirus coat protein. Desirably, the invention provides an isolated and purified nucleic acid encoding a chimeric adenovirus hexon protein as defined herein, wherein the nucleic acid sequence comprises a deletion of a region (or a plurality of such deletions) that encodes from about 1 to about 750 amino acids of the wild-type adenovirus coat protein, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the region deleted comprises a smaller region that encodes less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. In particular, optimally the deletion (e.g., of an adenoviral hexon protein) comprises the entirety of the 11 and/or 12 loop, or a sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Adl, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The invention also preferably provides an isolated and purified nucleic acid encoding a chimeric adenovirus hexon protein as defined herein, wherein the nucleic acid sequence comprises a deletion of one or more sequences selected from the group consisting of equivalents and conservatively modified variants of sequences that encode the entirety of the 11 and/or 12 loop, or SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID

NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Ad1, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

With respect to the nucleic acid sequence, an "equivalent" is a variation on the nucleic acid sequence such as can occur in different strains of adenovirus, and which either does or does not result in a variation at the amino acid level. Failure to result in variation at the amino acid level can be due, for instance, to degeneracy in the triplet code. A "conservatively modified variant" is a variation on the nucleic acid sequence that results in one or more conservative amino acid substitutions. In comparison, a "nonconservatively modified variant" is a variation on the nucleic acid sequence that results in one or more nonconservative amino acid substitutions.

In another preferred embodiment, the invention provides an isolated and purified nucleic acid encoding a chimeric adenovirus coat protein wherein the nucleic acid sequence further comprises a replacement of the deleted region (or a plurality of such replacements) with a spacer nucleic acid region (i.e., the nucleic acid sequence that encodes the aforementioned "spacer region") that encodes from about 1 to about 750 amino acids of the wild-type adenovirus coat protein, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the region deleted and replaced comprises a smaller region that encodes less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids.

Preferably, the spacer nucleic acid region comprises a FLAG octapeptide-encoding sequence [SEQ ID NO:49], and equivalents and conservatively modified variants of SEQ ID NO:49. Similarly, a spacer nucleic acid region can be employed that substitutes one or more coat protein encoding regions (particularly a hexon protein encoding region) of a particular adenoviral serotype with a coat protein encoding region (particularly a hexon protein encoding region) of another adenoviral serotype. Thus, preferably a spacer nucleic acid region present in a chimeric adenoviral hexon protein is selected from the group consisting of sequences that encode the entirety of the 11 and/or 12 loop, or SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Ad1, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra, and equivalents and conservatively modified variants of these sequences.

As described above with respect to the chimeric adenovirus coat protein, the spacer nucleic acid region (or a plurality thereof) simply can be incorporated into the coat protein in the absence of any deletions. These manipulations can be carried out so as to produce the above-described chimeric adenovirus coat protein.

The means of making such a chimeric adenoviral coat protein (i.e., by introducing conservative or nonconservative variations at either the level of DNA or protein) are known in the art, are described in the Examples which follow, and also can be accomplished by means of various commercially available kits and vectors

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(e.g., New England Biolabs, Inc., Beverly, MA; Clontech, Palo Alto, CA; Stratagene, LaJolla, CA, and the like). In particular, the ExSite™ PCR-based site-directed mutagenesis kit and the Chameleon™ double-stranded site-directed mutagenesis kit by Stratagene can be employed for introducing such mutations. Moreover, the means of assessing such mutations (e.g., in terms of effect on ability not to be neutralized by antibodies directed against wild-type hexon protein) are described in the Examples herein.

Accordingly, the present invention provides a preferred means of making a chimeric adenoviral coat protein, particularly a chimeric adenoviral hexon protein, which comprises obtaining an adenoviral genome encoding the wild-type adenovirus coat protein (e.g., the wild-type adenovirus hexon protein), and deleting one or more region(s) of the chimeric adenovirus coat protein (particularly the chimeric adenovirus hexon protein) comprising from about 1 to about 750 amino acids by modifying the corresponding nucleic acid coding sequence. Similarly, the invention provides a method of making a chimeric adenovirus coat protein (particularly a chimeric adenovirus hexon protein) which comprises obtaining an adenoviral genome encoding the wild-type adenovirus coat protein, deleting one or more region(s) of the adenovirus coat protein comprising from about 1 to about 750 amino acids by modifying the corresponding coding sequence, and replacing the deleted region(s) with a spacer region comprising from about 1 to about 300 amino acids by introducing a nucleic acid region (i.e., a "spacer nucleic acid region") that codes for same. Alternately, the spacer region preferably is simply incorporated into the coat protein (particularly the hexon protein) in the absence of any deletion. Optimally the spacer nucleic acid region encodes a nonconservative variation of the

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amino acid sequence of the wild-type adenovirus coat protein. The size of the DNA used to replace the native coat protein coding sequence may be constrained, for example, by impeded folding of the coat protein or improper assembly of the coat protein into a complex (e.g., penton base/hexon complex) or virion. DNA encoding 150 amino acids or less is particularly preferred for insertion/replacement in the chimeric coat protein gene sequence, and DNA encoding 50 amino acids or less is even more preferred.

Briefly, the method of mutagenesis comprises deleting one or more regions of an adenovirus coat protein, and/or inserting into an adenovirus coat protein one or more regions with a differing amino acid sequence, particularly by manipulating the DNA sequence. Several methods are available for carrying out such manipulations of adenovirus coat protein DNA sequences; these methods further can be used in combination. The method of choice depends on factors known to those skilled in the art, e.g., the size of the DNA region to be manipulated. For instance, convenient restriction sites (which further can be introduced into a sequence) can be used to introduce or remove segments of DNA, or entire genes or coding sequences. Alternately, other methods of mutagenesis involve the hybridization of a mismatched oligonucleotide to a region of single-stranded target DNA, extending the primer, for instance, using T7 DNA polymerase or other such means to produce a double-stranded heteroduplex, and isolating the mutant strand that incorporates the mismatched oligonucleotide from the parental nonmutant strand for use as a template and in further manipulations. The mutant strand can be separated from the parental strand using various selection means known to those skilled in the art (see, e.g., Kunkel et al., Methods Enzymol., 204, 125-139 (1991), as well as the underlying

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methodology employed in the $Chameleon^{TM}$ kit). Alternately, the parental strand can be selectively degraded, for instance, with use of enzymes that nick the nonmethylated strand of a hemi-methylated DNA molecule (e.g., HpaII, MspI, and Sau3AI), and by extending the mutant strand using 5-methyl-dCTP, which renders the strand resistant to cleavage by these enzymes. Along the same lines, an entirely PCR-based approach can be employed for making mutations (e.g., Kunkel, Proc. Natl. Acad. Sci., 82, 488-492 (1985); Costa et al., Nucleic Acids Res., 22, 2423 (1994)), for instance, such as the approach encompassed by the ExSiteTM kit. More generally, amino acid substitutions or deletions can be introduced during PCR by incorporating appropriate mismatches in one or both primers. Once the chimeric coat protein sequence has been produced, the nucleic acid fragment encoding the sequence further can be isolated, e.g., by PCR amplification using 5' and 3' primers, or through use of convenient restriction sites.

Vector Comprising a Chimeric Hexon Protein

A "vector" according to the invention is a vehicle for gene transfer as that term is understood by those skilled in the art, and includes viruses, plasmids, and the like. A preferred vector is an adenovirus, particularly a virus of the family Adenoviridae, and desirably of the genus Mastadenovirus (e.g., comprised of mammalian adenoviruses) or Aviadenovirus (e.g., comprised of avian adenoviruses). Such an adenovirus (or other viral vector) can be transferred by its own means of effecting cell entry (e.g., by receptor-mediated endocytosis), or can be transferred to a cell like a plasmid, i.e., in the form of its nucleic acid, for instance, by using liposomes to transfer the nucleic acid, or by microinjecting or transforming the DNA into the cell. The nucleic acid vectors that can be employed for

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gene transfer, particularly the adenoviral nucleic acid vectors, are referred to herein as "transfer vectors". Such nucleic acid vectors also include intermediary plasmid vectors that are employed, e.g., in the construction of adenoviral vectors.

Desirably an adenoviral vector is a serotype group C virus, preferably an Ad2 or Ad5 vector, although any other serotype adenoviral vector (e.g., group A including serotypes 12 and 31, group B including serotypes 3 and 7, group D including serotypes 8 and 30, group E including serotype 4, and group F including serotypes 40 and 41, and other Ad vectors previously described) can be employed. An adenoviral vector employed for gene transfer can be replication competent. Alternately, an adenoviral vector can comprise genetic material with at least one modification therein, which renders the virus replication deficient. The modification to the adenoviral genome can include, but is not limited to, addition of a DNA segment, rearrangement of a DNA segment, deletion of a DNA segment, replacement of a DNA segment, or introduction of a DNA lesion. A DNA segment can be as small as one nucleotide and as large as 36 kilobase pairs (i.e., the approximate size of the adenoviral genome) or, alternately, can equal the maximum amount which can be packaged into an adenoviral virion (i.e., about 38 kb). Preferred modifications to the group C adenoviral genome include modifications in the E1, E2, E3 and/or E4 regions. Similarly, an adenoviral vector can be a cointegrate, i.e., a ligation of adenoviral sequences with other sequences, such as other virus sequences, particularly baculovirus sequences, or plasmid sequences, e.g., so as to comprise a prokaryotic or eukaryotic expression vector.

In terms of an adenoviral vector (particularly a replication deficient adenoviral vector), such a vector can comprise either complete capsids (i.e., including a

viral genome such as an adenoviral genome) or empty capsids (i.e., in which a viral genome is lacking, or is degraded, e.g., by physical or chemical means). The capsid further can comprise nucleic acid linked to the surface by means known in the art (e.g., Curiel et al., Human Gene Therapy, 3, 147-154 (1992)) or can transfer non-linked nucleic acid, for instance, by adenoviral-mediated uptake of bystander nucleic acid (e.g., PCT International Application WO 95/21259).

Along the same lines, since methods are available for transferring an adenovirus in the form of its nucleic acid sequence (i.e., DNA), a vector (i.e., a transfer vector) similarly can comprise DNA, in the absence of any associated protein such as capsid protein, and in the absence of any envelope lipid. Inasmuch as techniques are available for making a RNA copy of DNA (e.g., in vitro transcription), and inasmuch as RNA viruses also can be employed as vectors or transfer vectors, a transfer vector also can comprise RNA. Thus, according to the invention whereas a vector comprises (and, further, may encode) a chimeric adenoviral coat protein, a transfer vector typically encodes a chimeric adenoviral coat protein (particularly a chimeric adenoviral hexon and/or fiber protein).

Based on this, the invention provides an adenoviral vector that comprises a chimeric coat protein (particularly a chimeric hexon and/or fiber protein) according to the invention. Preferably such a vector comprises a chimeric coat protein (particularly a chimeric adenovirus hexon protein and/or chimeric adenovirus fiber protein) as described above. Alternately, preferably the vector lacks wild-type fiber protein, e.g., the vector encodes a truncated or non-functional fiber protein, or fails to translate fiber protein. Such fiber mutations and the means of introducing fiber mutations are known to

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those skilled in the art (see, e.g., Falgout et al., \underline{J} . Virol., 62, 622-625 (1988)).

Of course, the chimeric adenoviral coat proteins include coat proteins in which the native (i.e., wild-type) hexon and/or fiber protein of an adenoviral vector is replaced by a hexon and or fiber amino acid sequence of a different adenoviral serotype such that the resultant adenoviral vector has a decreased ability or inability to be recognized by neutralizing antibodies directed against the corresponding wild-type coat protein. This replacement can comprise the entirety of the hexon and/or fiber amino acid sequence, or only a portion, as described above. Both proteins can be manipulated (e.g., in a single adenovirus), or only a single chimeric adenovirus coat protein can be employed, with the remaining coat proteins being wild-type.

A vector according to the invention (including a transfer vector) preferably comprises additional sequences and mutations, e.g., some that can occur within the coat protein coding sequence itself. In particular, a vector according to the invention further preferably comprises a nucleic acid encoding a passenger gene or passenger coding sequence. A "nucleic acid" is a polynucleotide (i.e., DNA or RNA). A "gene" is any nucleic acid sequence coding for a protein or an RNA molecule. Whereas a gene comprises coding sequences plus any non-coding sequences, a "coding sequence" does not include any non-coding (e.g., regulatory) DNA. A "passenger gene" or "passenger coding sequence" is any gene which is not typically present in and is subcloned into a vector (e.g., a transfer vector) according to the present invention, and which upon introduction into a host cell is accompanied by a discernible change in the intracellular environment (e.g., by an increased level of deoxyribonucleic acid (DNA), ribonucleic acid (RNA), peptide or protein, or by an

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"gene product" is either an as yet untranslated RNA
molecule transcribed from a given gene or coding sequence
(e.g., mRNA or antisense RNA) or the polypeptide chain
(i.e., protein or peptide) translated from the mRNA
molecule transcribed from the given gene or coding
sequence. A gene or coding sequence is "recombinant" if
the sequence of bases along the molecule has been altered
from the sequence in which the gene or coding sequence is
typically found in nature, or if the sequence of bases is
not typically found in nature. According to this
invention, a gene or coding sequence can be naturally
occurring or wholly or partially synthetically made, can
comprise genomic or complementary DNA (cDNA) sequences,
and can be provided in the form of either DNA or RNA.

Non-coding sequences or regulatory sequences include promoter sequences. A "promoter" is a DNA sequence that directs the binding of RNA polymerase and thereby promotes RNA synthesis. "Enhancers" are cis-acting elements of DNA that stimulate or inhibit transcription of adjacent genes. An enhancer that inhibits transcription is also termed a "silencer". Enhancers differ from DNA-binding sites for sequence-specific DNA binding proteins found only in the promoter (which also are termed "promoter elements") in that enhancers can function in either orientation, and over distances of up to several kilobase pairs, even from a position downstream of a transcribed region. According to the invention, a coding sequence is "operably linked" to a promoter (e.g., when both the coding sequence and the promoter constitute a passenger gene) when the promoter is capable of directing transcription of that coding sequence.

Accordingly, a "passenger gene" can be any gene, and desirably either is a therapeutic gene or a reporter gene. Preferably a passenger gene is capable of being expressed

in a cell in which the vector has been internalized. For instance, the passenger gene can comprise a reporter gene, or a nucleic acid sequence which encodes a protein that can be detected in a cell in some fashion. The passenger gene also can comprise a therapeutic gene, for instance, a therapeutic gene which exerts its effect at the level of RNA or protein. Similarly, a protein encoded by a transferred therapeutic gene can be employed in the treatment of an inherited disease, such as, e.g., the cystic fibrosis transmembrane conductance regulator cDNA for the treatment of cystic fibrosis. The protein encoded by the therapeutic gene can exert its therapeutic effect by resulting in cell killing. For instance, expression of the gene in itself may lead to cell killing, as with expression of the diphtheria toxin A gene, or the expression of the gene may render cells selectively sensitive to the killing action of certain drugs, e.g., expression of the HSV thymidine kinase gene renders cells sensitive to antiviral compounds including acyclovir, gancyclovir and FIAU (1-(2-deoxy-2-fluoro-b-Darabinofuranosil) -5-iodouracil). Moreover, the therapeutic gene can exert its effect at the level of RNA, for instance, by encoding an antisense message or ribozyme, by affecting splicing or 3' processing (e.g., polyadenylation), or by encoding a protein which acts by affecting the level of expression of another gene within the cell (i.e., where gene expression is broadly considered to include all steps from initiation of transcription through production of a processed protein), perhaps, among other things, by mediating an altered rate of mRNA accumulation, an alteration of mRNA transport, and/or a change in post-transcriptional regulation. Accordingly, the use of the term "therapeutic gene" is intended to encompass these and any other embodiments of that which is more commonly referred to as gene therapy

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and is known to those of skill in the art. Similarly, the recombinant adenovirus can be used for gene therapy or to study the effects of expression of the gene (e.g., a reporter gene) in a given cell or tissue in vitro or in vivo, or for diagnostic purposes.

Also, a passenger coding sequence can be employed in the vector. Such a coding sequence can be employed for a variety of purposes even though a functional gene product may not be translated from the vector sequence. For instance, the coding sequence can be used as a substrate for a recombination reaction, e.g., to recombine the sequence with the host cell genome or a vector resident in the cell. The coding sequence also can be an "anticoding sequence," e.g., as appropriate for antisense approaches. Other means of using the coding sequence will be known to one skilled in the art.

The present invention thus provides recombinant adenoviruses comprising a chimeric hexon protein and/or a chimeric fiber protein, and which preferably additionally comprise a passenger gene or genes capable of being expressed in a particular cell. The recombinant adenoviruses can be generated by use of a vector, specifically, a transfer vector, and preferably a viral (especially an adenoviral) or plasmid transfer vector, in accordance with the present invention. Such a transfer vector preferably comprises a chimeric adenoviral hexon and/or fiber gene sequence as previously described.

Similarly, the means of constructing such a transfer vector are known to those skilled in the art. For instance, a chimeric adenovirus coat protein gene sequence can simply be ligated into the vector using convenient restriction sites. Alternately, a wild-type adenovirus gene sequence can be mutagenized to create the chimeric coat protein sequence following its subcloning into a vector. Similarly, a chimeric coat protein gene sequence

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can be moved via standard molecular genetic techniques from a transfer vector into baculovirus or a suitable prokaryotic or eukaryotic expression vector (e.g., a viral or plasmid vector) for expression and evaluation of penton base binding, and other biochemical characteristics.

Accordingly, the present invention also provides recombinant baculoviral and prokaryotic and eukaryotic expression vectors comprising an aforementioned chimeric adenoviral coat protein gene sequence, which, along with the nucleic acid form of the adenoviral vector (i.e., an adenoviral transfer vector) are "transfer vectors" as defined herein. By moving the chimeric gene from an adenoviral vector to baculovirus or a prokaryotic or eukaryotic expression vector, high protein expression is achievable (approximately 5-50% of the total protein being the chimeric protein).

Similarly, adenoviral vectors (e.g., virions or virus particles) are produced using transfer vectors. For instance, an adenoviral vector comprising a chimeric coat protein according to the invention can be constructed by introducing into a cell, e.g., a 293 cell, a vector comprising sequences from the adenoviral left arm, and a vector comprising sequences from the adenoviral right arm, wherein there is a region of overlap between the sequences. As described in the Examples which follow, this methodology results in recombination between the sequences, generating a vector that comprises a portion of each of the vectors, particularly the region comprising the chimeric coat protein sequences.

The present invention thus preferably also provides a method of constructing an adenoviral vector that has a decreased ability or inability to be recognized by a neutralizing antibody directed against wild-type adenovirus hexon protein and/or fiber protein. This method comprises replacing a coat protein of the vector

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(i.e., a wild-type adenovirus hexon and/or fiber protein) with the corresponding chimeric adenovirus coat protein according to the invention to produce a recombinant adenoviral vector.

The coat protein chimera-containing particles are produced in standard cell lines, e.g., those currently used for adenoviral vectors. Deletion mutants lacking the fiber gene, or possessing shortened versions of the fiber protein, similarly can be employed in vector construction, e.g., H2d1802, H2d1807, H2d11021 (Falgout et al., supra), as can other fiber mutants. The fiberless particles have been shown to be stable and capable of binding and infecting cells (Falgout et al., supra).

Illustrative Uses and Benefits

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The present invention provides a chimeric coat protein that has a decreased ability or inability to be recognized by a neutralizing antibody directed against the corresponding wild-type coat protein, as well as vectors (including transfer vectors) comprising same. The chimeric coat protein (such as a chimeric hexon and/or fiber protein) has multiple uses, e.g., as a tool for studies in vitro of capsid structure and assembly, and capsomere binding to other proteins.

A vector (e.g., a transfer vector) comprising a chimeric coat protein can be used in strain generation, for instance, in generation of recombinant strains of adenovirus. Similarly, such a vector, particularly an adenoviral vector, can be used in gene therapy. Specifically, a vector of the present invention can be used to treat any one of a number of diseases by delivering to targeted cells corrective DNA, i.e., DNA encoding a function that is either absent or impaired, or a discrete killing agent, e.g., DNA encoding a cytotoxin that, for instance, is active only intracellularly.

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Diseases that are candidates for such treatment include, but are not limited to, cancer, e.g., melanoma, glioma or lung cancers; genetic disorders, e.g., cystic fibrosis, hemophilia or muscular dystrophy; pathogenic infections, e.g., human immunodeficiency virus, tuberculosis or hepatitis; heart disease, e.g., preventing restenosis following angioplasty or promoting angiogenesis to reperfuse necrotic tissue; and autoimmune disorders, e.g., Crohn's disease, colitis or rheumatoid arthritis. In particular, gene therapy can be carried out in the treatment of diseases, disorders, or conditions that require repeat administration of the corrective DNA and/or the adenoviral vector, and thus for which current adenoviral-mediated approaches to gene therapy are less than optimal.

Moreover, such a vector, particularly an adenoviral vector, can be used to deliver material to a cell not as a method of gene therapy, but for diagnostic or research purposes. In particular, a vector comprising a chimeric adenovirus coat protein according to the invention can be employed to deliver a gene either *in vitro* or *in vivo*, for research and/or diagnostic purposes.

For instance, instead of transferring a so-called therapeutic gene, a reporter gene or some type of marker gene can be transferred instead. Marker genes and reporter genes are of use, for instance, in cell differentiation and cell fate studies, as well as potentially for diagnostic purposes. Moreover, a standard reporter gene such as a β -galactosidase reporter gene, a gene encoding green fluorescent protein (GFP), or a β -glucuronidase gene can be used in vivo, e.g., as a means of assay in a living host, or, for instance, as a means of targeted cell ablation (see, e.g., Minden et al., BioTechniques, 20, 122-129 (1996); Youvan, Science, 268,

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264 (1995); U.S. Patent 5,432,081; Deonarain et al., <u>Br.</u>
<u>J. Cancer</u>, <u>70</u>, 786-794 (1994)).

Similarly, it may be desirable to transfer a gene to use a host essentially as a means of production in vivo of a particular protein. Along these lines, transgenic animals have been employed, for instance, for the production of recombinant polypeptides in the milk of transgenic bovine species (e.g., PCT International Application WO 93/25567). The use of an adenovirus according to the invention for gene transfer conducted for protein production in vivo further is advantageous in that such use should result in a reduced (if not absent) immune response as compared with the use of a wild-type adenovirus vector. Other "non-therapeutic" reasons for gene transfer include the study of human diseases using an animal model (e.g., use of transgenic mice and other transgenic animals including p53 tumor suppressor gene knockouts for tumorigenic studies, use of a transgenic model for impaired glucose tolerance and human Alzheimer's amyloid precursor protein models for the study of glucose metabolism and for the pathogenesis of Alzheimer's disease, respectively, etc.).

Furthermore, an adenoviral vector comprising a chimeric adenovirus coat protein and employed as described above is advantageous in that it can be isolated and purified by conventional means. For instance, it is likely that special cell lines will not need to be made in order to propagate adenoviruses comprising the chimeric coat proteins.

These aforementioned illustrative uses and recitation of benefits are by no means comprehensive, and it is intended that the present invention encompass such further uses which necessarily flow from, but are not explicitly recited, in the disclosure herein.

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Means of Administration

The vectors and transfer vectors of the present invention can be employed to contact cells either in vitro or in vivo. According to the invention "contacting" comprises any means by which a vector is introduced intracellularly; the method is not dependent on any particular means of introduction and is not to be so construed. Means of introduction are well known to those skilled in the art, and also are exemplified herein.

Accordingly, introduction can be effected, for instance, either in vitro (e.g., in an ex vivo type method of gene therapy or in tissue culture studies) or in vivo by methods that include, but are not limited to, electroporation, transformation, transduction, conjugation, triparental mating, (co-)transfection, (co-)infection, high velocity bombardment with DNA-coated microprojectiles, incubation with calcium phosphate-DNA precipitate, direct microinjection into single cells, and the like. Similarly, the vectors can be introduced by means of membrane fusion using cationic lipids, e.g., liposomes. Such liposomes are commercially available (e.g., Lipofectin®, Lipofectamine™, and the like, supplied by Life Technologies, Gibco BRL, Gaithersburg, MD). Moreover, liposomes having increased transfer capacity and/or reduced toxicity in vivo (see, e.g., PCT International Application WO 95/21259 and references reviewed therein) can be employed in the present invention. Other methods also are available and are known to those skilled in the art.

According to the invention, a "host" encompasses any host into which a vector of the invention can be introduced, and thus encompasses an animal, including, but not limited to, an amphibian, bird, insect, reptile, or mammal. Optimally a host is a mammal, for instance, a

rodent, primate (such as chimpanzee, monkey, ape, gorilla, orangutan, or gibbon), feline, canine, ungulate (such as ruminant or swine), as well as, in particular, a human.

Similarly, a "cell" encompasses any cell (or collection of cells) from a host into which an adenoviral. vector can be introduced, e.g., preferably an epithelial cell. Any suitable organs or tissues or component cells can be targeted for vector delivery. Preferably, the organs/tissues/cells employed are of the circulatory system (e.g., heart, blood vessels or blood), respiratory system (e.g., nose, pharynx, larynx, trachea, bronchi, bronchioles, lungs), gastrointestinal system (e.g., mouth, pharynx, esophagus, stomach, intestines, salivary glands, pancreas, liver, gallbladder), urinary system (e.g., kidneys, ureters, urinary bladder, urethra), nervous system (e.g. brain and spinal cord, or special sense organs such as the eye) and integumentary system (e.g., skin). Even more preferably the cells being targeted are selected from the group consisting of heart, blood vessel, lung, liver, gallbladder, urinary bladder, and eye cells.

Thus, the present invention preferably also provides a method of genetically modifying a cell. This method preferably comprises contacting a cell with a vector comprising a chimeric adenovirus hexon protein and/or a chimeric adenovirus fiber protein, wherein desirably the vector is an adenovirus vector. The method preferably results in the production of a host cell comprising a vector according to the invention.

Moreover, the method of the invention of genetically modifying a cell can be employed in gene therapy, or for administration for diagnosis or study. The application of this method in vivo optimally comprises administering to a patient in need of gene therapy (e.g., a patient suffering from a disease, condition or disorder) a therapeutically effective amount of a recombinant adenovirus vector

according to the invention. This method preferably can be employed as part of an ongoing gene therapy regimen, e.g., wherein the vector (e.g., a recombinant adenovirus vector) comprising the chimeric adenovirus coat protein is administered following (e.g., after from about 1 week to about 2 months) administration of a therapeutically effective amount of a vector comprising either the corresponding wild-type coat protein or a coat protein of a different adenoviral serotype. Alternately, the vector comprising the chimeric adenovirus coat protein can be employed as an initial attempt at gene delivery.

One skilled in the art will appreciate that suitable methods of administering a vector (particularly an adenoviral vector) of the present invention to an animal for purposes of gene therapy (see, for example, Rosenfeld et al. (1991), supra; Jaffe et al., Clin. Res., 39(2), 302A (1991); Rosenfeld et al., Clin. Res., 39(2), 311A (1991a); Berkner, supra), chemotherapy, vaccination, diagnosis, and/or further study are available. Although more than one route can be used for administration, a particular route can provide a more immediate and more For instance, effective reaction than another route. local or systemic delivery can be accomplished by administration comprising application or instillation of the formulation into body cavities, inhalation or insufflation of an aerosol, or by parenteral introduction, comprising intramuscular, intravenous, peritoneal, subcutaneous, intradermal, as well as topical administration. Clinical trials regarding use of gene therapy vectors in vivo are ongoing. The methodology employed for such clinical trials as well as further technologies known to those skilled in the art can be used to administer the vector of the present invention for the purpose of research, diagnosis and/or gene therapy.

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Pharmaceutically acceptable excipients also are well-known to those who are skilled in the art, and are readily available. The choice of excipient will be determined in part by the particular method used to administer the recombinant vector. Accordingly, there is a wide variety of suitable formulations for use in the context of the present invention. The following methods and excipients are merely exemplary and are in no way limiting.

Formulations suitable for oral administration can consist of (a) liquid solutions, such as an effective amount of the compound dissolved in diluents, such as water, saline, or orange juice; (b) capsules, sachets or tablets, each containing a predetermined amount of the active ingredient, as solids or granules; (c) suspensions in an appropriate liquid; and (d) suitable emulsions. Tablet forms can include one or more of lactose, mannitol, corn starch, potato starch, microcrystalline cellulose, acacia, gelatin, colloidal silicon dioxide, croscarmellose sodium, talc, magnesium stearate, stearic acid, and other excipients, colorants, diluents, buffering agents, moistening agents, preservatives, flavoring agents, and pharmacologically compatible excipients. Lozenge forms can comprise the active ingredient in a flavor, usually sucrose and acacia or tragacanth, as well as pastilles comprising the active ingredient in an inert base, such as gelatin and glycerin, or sucrose and acacia, emulsions, gels, and the like containing, in addition to the active ingredient, such excipients as are known in the art.

A vector of the present invention (including an adenoviral vector and a transfer vector), alone or in combination with other suitable components, can be made into aerosol formulations to be administered via inhalation. These aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like.

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They may also be formulated as pharmaceuticals for non-pressured preparations such as in a nebulizer or an atomizer.

Formulations suitable for parenteral administration include aqueous and non-aqueous, isotonic sterile injection solutions, which can contain anti-oxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. The formulations can be presented in unit-dose or multi-dose sealed containers, such as ampules and vials, and can be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid excipient, for example, water, for injections, immediately prior to use. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described.

Additionally, a vector of the present invention can be made into suppositories by mixing with a variety of bases such as emulsifying bases or water-soluble bases.

Formulations suitable for vaginal administration can be presented as pessaries, tampons, creams, gels, pastes, foams, or spray formulas containing, in addition to the active ingredient, such carriers as are known in the art to be appropriate.

The dose administered to an animal, particularly a human, in the context of the present invention will vary with the gene of interest, the composition employed, the method of administration, the particular site and organism undergoing administration, and the reason for the administration (e.g., gene therapy, diagnosis, means of producing a protein, further study, etc). Generally, the "effective amount" of the composition is such as to

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produce the desired effect in a host which can be monitored using several end-points known to those skilled in the art. For example, one desired effect might comprise effective nucleic acid transfer to a host cell. Such transfer can be monitored in terms of a therapeutic effect (e.g., alleviation of some symptom associated with the disease or syndrome being treated), or by further evidence of the transferred gene or coding sequence or its expression within the host (e.g., using the polymerase chain reaction, Northern or Southern hybridizations, or transcription assays to detect the nucleic acid in host cells, or using immunoblot analysis, antibody-mediated detection, or particularized assays to detect protein or polypeptide encoded by the transferred nucleic acid, or impacted in level or function due to such transfer). One such particularized assay described in the Examples which follow includes an assay for expression of a chloramphenicol acetyl transferase reporter gene.

Generally, to ensure effective transfer of the vectors of the present invention, it is preferable that from about 1 to about 5,000 copies of the vector be employed per cell to be contacted, based on an approximate number of cells to be contacted in view of the given route of administration. It is even more preferable that from about 1 to about 300 plaque forming units (pfu) enter each cell. However, this is just a general guideline which by no means precludes use of a higher or lower amount of a component, as might be warranted in a particular application, either in vitro or in vivo. For example, the actual dose and schedule can vary depending on whether the composition is administered in combination with other pharmaceutical compositions, or depending on interindividual differences in pharmacokinetics, drug disposition, and metabolism. Similarly, amounts can vary in in vitro applications depending on the particular cell

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type utilized or the means by which the vector is transferred. One skilled in the art easily can make any necessary adjustments in accordance with the necessities of the particular situation.

The following examples further illustrate the present invention and, of course, should not be construed as in any way limiting its scope.

Example 1

This example describes experiments investigating adenoviral anti-vector neutralizing immunity.

To clarify the phenomenon of neutralizing immunity, an animal having circulating antibodies to one adenoviral vector type received intratracheal administration of another serotype adenoviral vector, and gene expression commanded by the second vector was monitored.

Specifically, either an Ad4 or Ad5 wild-type vector was administered to the lungs of Sprague-Dawley rats. Ten days later, an Ad5 reporter vector was administered to the lungs of the same animals. This reporter vector, which is referred to herein as the "pure 5" vector, comprises an E1-E3- type 5 adenoviral vector which expresses the chloramphenical acetyl transferase (CAT) gene driven by the cytomegalovirus early/intermediate promoter/enhancer (CMV) (i.e., AdCMVCATgD described in Kass-Eisler et al., Proc. Natl. Acad. Sci., 15, 11498-11502 (1993)).

About twenty-four hours following administration of the "pure 5" vector, CAT activity was measured in homogenized lung tissue using a CAT assay as previously described (Kass Eisler et al. (1993), supra). CAT activity was monitored at various times thereafter up to 10 days following introduction of the "pure 5" vector. CAT activity was determined relative to the "pure 5" vector administered to naive animals (i.e., expression measured under this condition was considered 100%). The

results of these studies are set out in **Table 1**, and are further reported in Mastrangeli et al., <u>Human Gene</u>

<u>Therapy</u>, <u>1</u>, 79-87 (1996).

Table 1. Effect of anti-serotype 4 (group E)
neutralizing antibodies on the ability of a "pure 5"
adenoviral vector to transfer a CAT reporter gene to
the lung

Time (0 hours)	Time (10 days)	CAT Activity
		0%
	pure 5	100%
Ad5	pure 5	0%
Ad4	pure 5	105±10%

These results confirm that in the presence of neutralizing antibodies elicited against one adenoviral group (e.g., against group E, serotype 4), it is possible to efficiently transfer and express a gene in vivo using an adenoviral vector derived from another group (e.g., derived from group C, serotype 5). Neutralizing immunity evoked against one serotype group does not protect against infection by another group of adenovirus. These data support the paradigm of alternating adenoviral vectors derived from different subgroups as a strategy to circumvent anti-adenoviral humoral immunity.

Example 2

The predominant epitopes that evoke neutralizing immunity are located on the fiber and hexon, but mainly on hexon. Based on this, the effect of switching the fiber protein was investigated. A vector was constructed that was identical to the "pure 5" vector except that the fiber gene was switched from a serotype 5, group C fiber to a

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serotype 7, group B fiber. The resultant vector is referred to herein as the "5 base/7 fiber" vector.

The Ad5/Ad7 fiber construct was generated as shown in Figure 1. An approximately 2.7 kb (Ad5 28689-31317 bp) fragment in pAd70-100 was replaced with a PacI linker (pAd70-100dlE3.Pac). A BamHI linker was inserted at a MunI site as indicated in Figure 2 to produce pAd70-100dlE3.Pac.Bam. A PCR-amplified PacI-BamHI fragment of approximately 1.1 kb containing the Ad7 fiber gene was inserted into pAd70-100dlE3.Pac.Bam to produce pAd70-100dlE3.fiber7.

In order to assess the ability of the Ad5 virus with Ad7 fiber to infect cells in vitro and in vivo, reporter gene assays were performed. A replication-defective recombinant adenoviral reporter vector designated AdCMV-CATNeo was used in the reporter gene assay. The reporter vector consists of the adenoviral origin of replication and viral packaging sequences, a combination of strong eukaryotic promoter (cytomegalovirus or CMV-1) and splicing elements, the bacterial chloramphenical acetyl transferase (CAT) gene sequence, the mouse $\beta^{\text{maj}}\text{-globin}$ poly(A) site, the neomycin gene sequence (Neo), and sufficient adenoviral DNA to allow for overlap recombination.

The reporter vector was used to generate AdCMV-CATNeo, AdCMV-CATNeo-dlE3 (AdCMV-CATNeo + pAd70-100dlE3) and AdCMV-CATNeo-dlE3-Fiber7 (AdCMV-CATNeo + pAd70-1001E3.Fiber7) viruses. Each virus was grown in large scale, i.e., a one liter suspension of human embryonic kidney 293 cells, to yield virus at a concentration of 10¹² particles/ml. A549 cells were infected with an estimated 100, 300 or 1,000 particles/cell of one of the three viruses. After 48 hours, the cells were harvested and lysates were prepared as described in Kass-Eisler et al.

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(1993), supra. Using 50 μ l of each lysate, CAT assays were performed and acetylated chloramphenical products were separated by thin layer chromatography using chloroform:methanol (95:5). The results of the assays confirm that each virus was able to infect cells and express gene products at appropriate levels. Accordingly, the virus in which the native fiber was replaced with a nonnative fiber could infect cells and express genes like the parental virus.

Following this study, adult Sprague-Dawley rats were infected with 108 viral particles by direct cardiac injection as described in Kass-Eisler et al. (1993), supra. Five days later, the rats were sacrificed, cardiac lysates were prepared, and CAT assays were performed. amount of the CAT gene product produced was compared between the dlE3 and dlE3-Fiber7 viruses. Results indicated that both viruses were able to infect cells in vivo. The replacement of the wild-type Ad5 fiber gene with that of Ad7 did not impair the ability of the virus to infect cells. Accordingly, the virus in which the native fiber was replaced with a nonnative fiber could also infect cells and express genes like the parental virus in vivo. These results support the utility of adenovirus with chimeric fiber in the context of gene therapy.

Example 3

This example describes the effect on neutralizing immunity of switching the fiber protein of an adenovirus from one serotype to another.

The "pure 5" and "5 base/7 fiber" vectors described in the preceding Example were administered to Sprague-Dawley rats which either were naive or pre-immunized against wild-type Ad5. For these experiments, wild-type Ad5 or wild-type Ad7 (6 x 10 particles in phosphate

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buffered saline (PBS)) was administered intraperitoneally as a primary inoculation. Seventeen days later, serum samples were taken, and about 6 x 10⁹ particles in about 50 µl of PBS was injected. At about 120 hours following injection the animals were sacrificed, serum and heart tissue were harvested, and heart tissue was processed for CAT assays as previously described (Kass-Eisler et al. (1993), supra). CAT assays also were performed on heart lysates of rat hearts infected with the "pure 5" vector or "5 base/7 fiber" vector alone.

Administration of either vector to naive animals resulted in comparable levels of CAT in heart tissue. In comparison, administration of either the "pure 5" vector or the "5 base/7 fiber" vector to the animals that were pre-immunized against the "pure 5" vector resulted in a reduction of CAT levels by more than two orders of magnitude as compared with mock-infected controls. These and further results are reported in Gall et al., <u>J.</u> Virol., 70, 2116-2163 (1996).

These results confirm that switching the fiber from that of adenoviral serotype 5 group C vector to that of an adenoviral serotype 7 group B vector by itself is insufficient to allow the vector to escape neutralizing antibodies generated against an adenoviral vector comprising Ad5 fiber. These results imply that antibodies against adenoviral structures other than fiber also are important in the process of neutralizing immunity. Furthermore, whereas switching the fiber serotype to another serotype may be insufficient in and of itself to allow an adenovirus to escape immune detection, such switching when done in combination with removal of other epitopes may be desirable, for instance, to reduce an immune response.

Example 4

This example describes the construction of adenovirus vectors wherein the neutralizing immunity-evoking epitopes have been modified. In particular, this example describes vectors comprising chimeric adenoviral hexon protein, wherein the hexon neutralizing immunity-evoking epitopes are modified.

The results of the prior example indicate that it is possible to develop vectors for repeat administration in gene therapy from non-group C adenovirus, thus circumventing pre-existing neutralizing immunity. As another strategy, the dominant neutralizing immunity-evoking epitopes on existing group C vectors can be modified to render the vectors less susceptible (or "stealth") to the existing neutralizing immunity. For instance, adenoviral type 5-based E1 E3 CAT-expressing vectors can be constructed that have the same genetic composition as the "pure 5" and "5 base/7 fiber" vectors described above, except for possessing a gene encoding a chimeric hexon that is not recognized by pre-existing anti-type 5 neutralizing immunity.

To derive the vectors, the chimeric hexon gene present in the "pure 5" parental vector can be modified, in particular, 11 and/or 12 can be altered. The hexon modifications that can be made on the "pure 5" CAT vector, or other adenoviral vector (such as any other adenoviral serotype vector), include, but are not limited to: (1) hexon with 11 deleted in its entirety; (2) hexon with 12 deleted in its entirety; (3) hexon with both 11 and 12 deleted; (4) hexon with any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7, deleted; (5)-(8) hexon with a FLAG octamer epitope (i.e., Asp Tyr Lys Asp Asp Asp Asp Lys [SEQ ID NO:50]; Hopp et al., Biotechnology, 6, 1205-1210 (1988)) substituted for 11, 12, or both 11 and 12, or any one or more of HVR1, HVR2, HVR3, HVR4, HVR5,

HVR6 or HVR7; (9)-(12) hexon with a FLAG octamer epitope [SEQ ID NO:50] inserted into 11, 12, or both 11 and 12; (13)-(16) hexon with comparable epitopes from Ad7 (group B) (GenBank® Data Bank Accession Number x76551 for Ad7 hexon, and Number M73260 for Ad5 hexon) or Ad2, or any other adenoviral serotype, substituted for 11, 12, both 11 and 12, respectively, or for any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7; (17)-(20) hexon with comparable epitopes from Ad7 (group B) (GenBank® Data Bank Accession Number x76551 for Ad7 hexon, and Number M73260 for Ad5 hexon) or Ad2, or any other adenoviral serotype, inserted into 11, 12, both 11 and 12, respectively, or any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7; and (21) complete substitution of the hexon from Ad2 or another adenoviral serotype, for the Ad5 hexon. The use of the FLAG octamer epitope provides a sequence for incorporation in the chimeric hexon protein that is different from the Ad5 hexon loop sequences, and also provides a positive control using available specific anti-FLAG antibodies (Hopp et al., supra).

These chimeric hexon proteins (and vectors containing them) can be made in several steps. To modify the hexon in the "pure 5" vector, a viral or plasmid vector can be constructed to contain the hexon type 5 coding sequence in a cassette that can be easily modified. The hexon is read off the 1 strand of the L3 transcription unit, i.e., map units 51.6 to 59.7, comprising a region of about 2.9 kb. The two other transcripts that also are encoded by L3 — i.e., polypeptide VI and a 23 kDa protein — do not overlap the hexon coding sequence. Moreover, there are no other coding sequences on the r strand that would be altered by the modification of the hexon coding sequence.

Thus, all the modifications of the type 5 hexon can be made using a "hexon 5 cassette" comprised of an

approximate 6.7 kb SfiI-SfiI fragment of the "pure 5" CAT vector. SfiI cuts Ad5 into 3 fragments, the center 6.7 kb fragment (i.e., comprising about 16,282 to 22,992 base pairs, as identified by agarose gel electrophoresis) of which contains all of the L3 region plus some overlap. The "hexon 5 cassette" can be subcloned into a commercially available vector having restriction sites and the like making the vector easily manipulable in terms of modification and recovery of subcloned sequences. One such vector appropriate for subcloning is either the SK or KS version of the pBlueScript® phagemid (Stratagene, LaJolla, CA).

The "hexon 5 cassette" can be mutagenized to generate site-specific mutations in the cloned DNA segment. Several methods are available for carrying out sitespecific mutagenesis. The 11 and 12 deletions, insertions, or replacements (or deletions, insertions, or replacements in HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7 regions contained therein) can be made by deleting the relevant sequences using restriction enzymes that cut uniquely within the vector inserts, or other similar means, e.g., by ligating in an end-polished, or otherwise modified, PCR product. Alternately, the hexon sequence contained in the hexon 5 cassette can be modified, e.g., using single-stranded mutagenesis in M13mp8 or some other convenient vector, and using appropriate oligonucleotides encompassing the flanking sequences for identification of plaques as described by Crompton et al., supra. Alternately, a commercially available kit such as the $\operatorname{ExSite}^{\operatorname{TM}}$ PCR-based site-directed mutagenesis kit and the Chameleon™ double-stranded site-directed mutagenesis kit by Stratagene can be used to introduce insertions, point mutations, or deletions into the chimeric hexon sequence without any need for subcloning into an M13, or other special vector.

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Similarly, the FLAG octapeptide sequence (Hopp et al., supra) can be introduced into the vectors (i.e., in the presence or absence of any deletion) by inserting the relevant 24 base pair sequence (GAY TAY AAR GAY GAY GAY AAR [SEQ ID NO:50], wherein Y is C or T/U, and R is A or G)). The replacement of Ad5 hexon loop epitopes with comparable sequences of Ad7, Ad2, or any other adenoviral serotype, or an incorporation of these sequences in the absence of any deletion, can be accomplished by using unique restriction sites, or using one of the aforementioned means of mutagenesis. This usefully creates new serotypes of adenoviral vectors. For example, The replacement of the wildtype hexon protein of Ad5 with the chimeric Ad5 hexon comprising Ad7 hexon loops 1 and 2 gives rise to an adenoviral vector that is effectively neutralized by Ad7 neutralizing antibodies (i.e., neutralizing antibodies raised in response to Ad7 innoculation of a naïve animal), but not by Ad5 neutralizing antibodies.

Moreover, both hypervariable loops 1 and 2 can be deleted from a serotype 5 or another serotype adenoviral vector. Adenoviral vectors and there genomes comprising these deletions are useful as a starting point to create other adenoviral vectors having loop replacements, as a tool for studying hexon structure-function relationships, and under some circumstances as a gene transfer vector with limited vulnerability to the adaptive immune system.

Example 5

This example describes the method of replacing the hexon protein of one serotype adenoviral vector with the hexon protein of another serotype adenoviral vector to generate a recombinant adenovirus. As representative of this method, the hexon protein of an Ad5 vector was replaced with the hexon protein of an Ad2 vector. This

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example also describes the method of incorporating the chimeric hexon proteins of the preceding Example into a vector to make a recombinant adenovirus.

Using standard molecular biology techniques, the Ad5 hexon gene open reading frame (ORF) was replaced with the Ad2 hexon gene ORF in such a fashion so as to maintain the proper Ad5 sequences upstream and downstream of the hexon gene. Adenoviral vectors comprising modified or chimeric hexon proteins can be constructed by homologous recombination using standard techniques and human embryonic kidney 293 cells (see, e.g., Rosenfeld et al. (1991), supra; Rosenfeld et al. (1992), supra). For instance, map units 0 to 57.3 of dlAd5NCAT (Gall et al., supra) can be isolated by Bsu36I digestion, and map units 58.4 to 100 of dlAd5NCAT can be isolated by DrdI digestion. These DNA fragments can be transfected into 293 cells along with pH5-2.

A neutralizing antibody directed against the parental vector can be employed to facilitate the generation of hexon replacement constructs. For example, when replacing the loop 1 and loop 2 regions of an Ad5 vector with Ad7 loop sequences, anti-Ad5 neutralizing polyclonal or monoclonal antibodies (directed against the loops 1 and 2 of Ad5 hexon) can be added to a the medium of cells in which the chimeric vector is being propagated. The presence of the Ad5 neutralizing antibodies substantially blocks the propagation of the undesired wildtype Ad5 vector(s), while the chimeric vector is unaffected. Furthermore, the recombinant vectors comprising a chimeric hexon ORF can be generated by homologous recombination using a plasmid that carries a marker gene, such as Green Fluorescent Protein (GFP), adjacent to the chimeric or novel hexon ORF (e.g., between the fiber and hexon genes). In this way, genomes that could harbor the chimeric hexon gene should also harbor the marker gene. The marker gene

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would then be expressed as a late protein, so that cells that potentially comprise the desired adenoviral genome can be easily identified.

Similarly, vectors (particularly adenoviral vectors) can be constructed that have the aforementioned hexon modifications, and which have further modifications, for instance, in the adenoviral fiber coding sequences. This can be accomplished by making the hexon modifications described above, and using different parental plasmids for homologous recombination, such as parental plasmids comprising mutations in fiber coding sequences. In particular, the "5 base/7 fiber" vector can be employed as a starting vector for vector construction.

All of the viral vectors prepared according to this example can be plaque-purified, amplified, and further purified using standard methods (Rosenfeld et al. (1991), supra; Rosenfeld et al. (1992), supra).

Example 6

This example describes a characterization of the activity in vitro and in vivo of the vectors described in the preceding Examples.

Each of the viruses prepared as described in the preceding Examples can be evaluated in vitro and in vivo using standard methods as previously described (e.g., Kass-Eisler et al., supra), and as set forth herein. In particular, for the in vitro studies, the various vectors along with control vectors (e.g., the "pure 5" and "5 base/7 fiber" vectors, and the Ad5 wild-type vector) can be added to human lung carcinoma A549 cells alone, or in the presence of dilutions of serum from hosts infected with Ad5, Ad7, "pure 5" CAT vector, or "5 base/7 fiber" CAT vector, or anti-FLAG epitope serum. The cells are then evaluated for CAT activity to determine the ability

of antibodies present in the serum to block gene expression.

The in vivo studies can be carried out in Sprague-Dawley rats. The Sprague-Dawley rat as opposed to the mouse or cotton rat is preferred for these experiments since the rat is non-permissive, and the wild-type adenovirus cannot replicate in this host. Accordingly, immunizations can be carried out using wild-type viruses (e.g., wild-type Ad5 or Ad7), the "pure 5" CAT vector, and the "5 base/7 fiber" CAT vector by intravenous administration (e.g., Kass-Eisler et al., supra). At various times ranging from about one to about four weeks later, the vector of interest can be administered intravenously or directly into the airways of the host. Whereas intravenous administration allows an assessment of the "worst case scenario" (i.e., wherein the vector is in immediate contact with the circulating humoral immune system, and thus the strongest immune response is to be expected), introduction in the airways of the host allows an evaluation of a compartmentalized and mucosal humoral immune response.

CAT activity can be quantified as previously described in all the relevant organs, e.g., liver, heart, and lung for intravenous administration, and lung only for respiratory administration. Appropriate standards can be used to compensate for variations in organ expression of CAT activity (see e.g., Kass-Eisler et al., Gene Therapy, 2 395-402 (1994)). The in vitro and in vivo results can be compared and assessed using standard statistical methods.

All of the references cited herein, including the GenBank® Data Bank sequence information, are hereby incorporated in their entireties by reference.

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While this invention has been described with emphasis upon preferred embodiments, it will be obvious to those of ordinary skill in the art that the preferred embodiments can be varied. It is intended that the invention can be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the appended claims.

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 - (F) POSTAL CODE (ZIP): 22043
 - (ii) TITLE OF INVENTION: CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME
- (iii) NUMBER OF SEQUENCES: 56
- (iv) COMPUTER READABLE FORM:

528

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145

(A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO) (vi) PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: US 8-816346 (B) FILING DATE: 13-MAR-1997 INFORMATION FOR SEQ ID NO:1: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2907 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1: ATG GCT ACC CCT TCG ATG ATG CCG CAG TGG TCT TAC ATG CAC ATC TCG 48 Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser 96 GGC CAG GAC GCC TCG GAG TAC CTG AGC CCC GGG CTG GTG CAG TTT GCC Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala 20 25 144 CGC GCC ACC GAG ACG TAC TTC AGC CTG AAT AAC AAG TTT AGA AAC CCC Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro 35 192 ACG GTG GCA CCT ACG CAC GAC GTA ACC ACA GAC CGG TCC CAG CGT TTG Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu 50 55 240 ACG CTG CGG TTC ATC CCT GTG GAC CGC GAG GAT ACC GCG TAC TCG TAC Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr AAA GCG CGG TTC ACC CTG GCT GTG GGT GAC AAC CGT GTG CTT GAT ATG 288 Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met . 85 336 GCT TCC ACG TAC TTT GAC ATC CGC GGC GTG CTG GAC AGG GGG CCT ACT Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr 110 100 105 384 TTT AAG CCC TAC TCC GGC ACT GCC TAC AAC GCT CTA GCT CCC AAG GGC Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly 120 125 115 GCT CCT AAC TCC TGT GAG TGG GAA CAA ACC GAA GAT AGC GGC CGG GCA 432 Ala Pro Asn Ser Cys Glu Trp Glu Gln Thr Glu Asp Ser Gly Arg Ala 140 130 135 480 Val Ala Glu Asp Glu Glu Glu Glu Asp Glu Glu Glu Glu Glu 160 150 155

GAA GAG CAA AAC GCT CGA GAT CAG GCT ACT AAG AAA ACA CAT GTC TAT

Glu Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr

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GCC Ala	CAG Glr	GC Al	a F	CT Pro	TTG Leu	TCT Ser	GGA Gly	GAA Glu	ACA Thr 185	ATT	AC Th	A A	AA ys	AGC Ser	GGG Gly 190		A C	AA ln	576	
ATA Ile	GGF Gly	TC Se 19	er P	SAC Asp	AAT Asn	GCA Ala	GAA Glu	ACA Thr 200	CAA Gln	GCT	AA Ly	AA C	CT Pro	GTA Val 205	TAC	GC.	A G a A	AT .sp	624	
CCT	TC0 Ser	c T	AT (CAA	CCA Pro	GAA Glu	CCT Pro 215	CAA Gln	ATT Ile	GGC Gly	C GI y Gi	Lu	CT Ser 220	CAG Gln	TGG	AA As	C G	AA Slu	. 672	
GCT Ala	As	r G(p A)	CT l	AAT Asn	GCG Ala	GCA Ala 230	GGA Gly	GGG Gly	AGA Arg	GT(רו ד	TT 1 eu 1 35	AAA Lys	AAA Lys	ACF Thr	A AC	T C	CCC Pro 240	720	
		A CO	CA '	TGC Cys	TAT Tyr 245	Gly	TCT Ser	TAT Tyr	GCC	AG Ar 25	g r	CT i	ACA Thr	AAT Asn	CCT	TT Ph 25	•	GGT Gly	768	
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GT Va	T GA l As	p L	TG eu 75	CAA Gln	TTC	TTC Phe	TCA Ser	AAT Asr 280	1 111	r AC	C T	CT	TTG Leu	AAC Asi 28	•	C CC p A:	GG rg	CAA Gln	864	
GG G1	y As	AT G	CT	ACT Thr	AAA Lys	A CCA	A AAA Lys	s val	GT' L Va	r TI l Le	rG T	TAC Tyr	AGT Ser 300	G L	A GA u As	T G p V	TA al	AAT Asn	912	!
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		AT :	TCT Ser	AAA Lys	A GC's Ala	a Me	G TT t Le	G GG u Gl	r CA y Gl	n G	AA ln 30	TCT Ser	AT(G CC	A AF	,,,	GA Arg 335	CCC Pro	·1008	3
Al A:	AT T sn T	AC I	ATT Ile	GC: Ala 340	a Ph	C AG e Ar	G GA g As	C AA p As	T TI n Ph 34	ie T	TT le	GGC Gly	CT.	A Al u Me	, L I	AT I yr I	TAT Tyr	AAC Asn	. 105	6
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		GCT Ala	GT Va	A GA 1 As	sp Se	CC 111	AT G	AT C	CA G	sp '	GTT Val 410	AL	A AT	rc A le I	TT (SAA Glu	AAC Asi 415	•	r 124	18
(GGA Gly	ACT Thr	GA Gl	u As	л С	ייז איני	TG C eu P	CA A ro A	sn 1	AT Yr 125	TGT Cys	TT Ph	T Co	CT C	י ששנ	GGG Gly 430	GG'	r AT y Il	T 12'	96

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								Asn			TCA Ser	. •	1344
			GAT Asp										1392
			GGA Gly										1440
			TGG Trp 485										1488
			CTA Leu										1536
			TAC Tyr	-									1584
			TAC Tyr						Ser				1632
			AAT Asn										1680
			TTG Leu 565								ATT Ile		1728
			AAG Lys				Lys						1776
			TAT Tyr										1824
	_		CTG Leu	_									1872
			ATT Ile										1920
			ACG Thr 645										1968
	•		GAC Asp			_					ATA Ile		2016
										Arg	 TGG Trp		2064

GCA GCA TTT CGC GGT TGG GCC TTC ACA CGC TTG AAG ACA AAG GAA ACC Ala Ala Phe Arg Gly Trp Ala Phe Thr Arg Leu Lys Thr Lys Glu Thr	2112
690 CCT TCC CTG GGA TCA GGC TAC GAC CCT TAC TAC ACC TAC TCT GGC TCC Pro Ser Leu Gly Ser Gly Tyr Asp Pro Tyr Tyr Thr Tyr Ser Gly Ser 700 710	2160
ATA CCA TAC CTT GAC GGA ACC TTC TAT CTT AAT CAC ACC TTT AAG AAG Ile Pro Tyr Leu Asp Gly Thr Phe Tyr Leu Asn His Thr Phe Lys Lys 725	2208
GTG GCC ATT ACC TTT GAC TCT TCT GTT AGC TGG CCG GGC AAC GAC CGC Val Ala Ile Thr Phe Asp Ser Ser Val Ser Trp Pro Gly Asn Asp Arg 740	2256
CTG CTT ACT CCC AAT GAG TTT GAG ATT AAA CGC TCA GTT GAC GGG GAG Leu Leu Thr Pro Asn Glu Phe Glu Ile Lys Arg Ser Val Asp Gly Glu 755	2304
GGC TAC AAC GTA GCT CAG TGC AAC ATG ACC AAG GAC TGG TTC CTG GTG GIY Tyr Asn Val Ala Gln Cys Asn Met Thr Lys Asp Trp Phe Leu Val	2352
CAG ATG TTG GCC AAC TAC AAT ATT GGC TAC CAG GGC TTC TAC ATT CCA Gln Met Leu Ala Asn Tyr Asn Ile Gly Tyr Gln Gly Phe Tyr Ile Pro 795	2400
GAA AGC TAC AAG GAC CGC ATG TAC TCG TTC TTC AGA AAC TTC CAG CCC Glu Ser Tyr Lys Asp Arg Met Tyr Ser Phe Phe Arg Asn Phe Gln Pro 815	2448
ATG AGC CGG CAA GTG GTT GAC GAT ACT AAA TAC AAG GAG TAT CAG CAG Met Ser Arg Gln Val Val Asp Asp Thr Lys Tyr Lys Glu Tyr Gln Gln 820 825	2496
GTT GGA ATT CTT CAC CAG CAT AAC AAC TCA GGA TTC GTA GGC TAC CTC Val Gly Ile Leu His Gln His Asn Asn Ser Gly Phe Val Gly Tyr Leu 835	2544
GCT CCC ACC ATG CGC GAG GGA CAG GCT TAC CCC GCC AAC GTG CCC TAC Ala Pro Thr Met Arg Glu Gly Gln Ala Tyr Pro Ala Asn Val Pro Tyr	2592
CCA CTA ATA GGC AAA ACC GCG GTT GAC AGT ATT ACC CAG AAA AAG TTT Pro Leu Ile Gly Lys Thr Ala Val Asp Ser Ile Thr Gln Lys Lys Phe 875	2640
CTT TGC GAT CGC ACC CTT TGG CGC ATC CCA TTC TCC AGT AAC TTT ATG Leu Cys Asp Arg Thr Leu Trp Arg Ile Pro Phe Ser Ser Asn Phe Met 895	2688
TCC ATG GGC GCA CTC ACA GAC CTG GGC CAA AAC CTT CTC TAC GCC AAC Ser Met Gly Ala Leu Thr Asp Leu Gly Gln Asn Leu Leu Tyr Ala Asn	2736
TCC GCC CAC GCG CTA GAC ATG ACT TTT GAG GTG GAT CCC ATG GAC GAG Ser Ala His Ala Leu Asp Met Thr Phe Glu Val Asp Pro Met Asp Glu 925	2784
CCC ACC CTT CTT TAT GTT TTG TTT GAA GTC TTT GAC GTG GTC CGT GTG Pro Thr Leu Leu Tyr Val Leu Phe Glu Val Phe Asp Val Val Arg Val 930 935	2832

63

2880 CAC CAG CCG CAC CGC GGC GTC ATC GAG ACC GTG TAC CTG CGC ACG CCC His Gln Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro 955 960 945 950 2907 TTC TCG GCC GGC AAC GCC ACA ACA TAA Phe Ser Ala Gly Asn Ala Thr Thr 965 (2) INFORMATION FOR SEQ ID NO:2: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 968 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser 15 Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala .25 Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu 50 Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr 65 Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr 105 Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly 120 • 125 115 · Ala Pro Asn Ser Cys Glu Trp Glu Gln Thr Glu Asp Ser Gly Arg Ala 130 135 Val Ala Glu Asp Glu Glu Glu Glu Asp Glu Asp Glu Glu Glu Glu Glu 150 145 155 Glu Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr 165 170 Ala Gln Ala Pro Leu Ser Gly Glu Thr Ile Thr Lys Ser Gly Leu Gln 185 180 Ile Gly Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val Tyr Ala Asp 200 205 195 Pro Ser Tyr Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp Asn Glu 210 . 215 Ala Asp Ala Asn Ala Ala Gly Gly Arg Val Leu Lys Lys Thr Thr Pro

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Met Lys Pro Cys Tyr Gly Ser Tyr Ala Arg Pro Thr Asn Pro Phe Gly Gly Gln Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys Val Asp Leu Gln Phe Phe Ser Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro Lys Val Val Leu Tyr Ser Glu Asp Val Asn Met Glu Thr Pro Asp Thr His Leu Ser Tyr Lys Pro Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly Gln Gln Ser Met Pro Asn Arg Pro Asn Tyr Ile Ala Phe Arg Asp Asn Phe Ile Gly Leu Met Tyr Tyr Asn Ser Thr Gly Asn Met Gly Val Leu Ala Gly Gln Ala Ser Gln Leu Asn Ala Val Val Asp Leu Gln Asp Arg Asn Thr Glu Leu Ser Tyr Gln Leu Leu Leu Asp Ser Ile Gly Asp Arg Thr Arg Tyr Phe Ser Met Trp Asn Gln Ala Val Asp Ser Tyr Asp Pro Asp Val Arg Ile Ile Glu Asn His Gly Thr Glu Asp Glu Leu Pro Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile

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Gln	Val	Pro	Gln 580	Lys	Phe	Phe	Ala	Ile 585	Lys	Asn	Leu	Leu	Leu 590	Leu	Pro
Gly	Ser	Tyr 595	Thr	Tyr	Glu	Trp	Asn 600	Phe.	Arg	Lys	Asp	.Val 605	Asn	Met	Val
Leu	Gln 610	Ser	Ser	Leu	Gly	Asn 615	Asp	Leu	Arg	Val	Asp 620	Gly	Ala	Ser	Ile
Lys 625	Phe	Asp	Ser	Ile	Cys 630	Leu	Tyr	Ala	Thr	Phe 635	Phe	Pro	Met	Ala	His 640
Asn	Thr	Ala	Ser	Thr 645	Leu	Glu	Ala	Met	Leu 650	Arg	Asn	Asp	Thr	Asn 655	Asp
Gln	Ser	Phe	Asn 660	Asp	Tyr	Leu	Ser	Ala 665	Ala	Asn	Met	Leu	Tyr 670	Pro.	Ile
Pro	Ala	Asn 675	Ala	Thr	Asn	Val	Pro 680	Ilė	Ser	Ile	Pro	Ser 685	Arg	Asn	Trp
Ala	Ala 690	Phe	Arg	Gly	Trp	Ala 695	Phe	Thr	Arg	Leu	Lys 700	Thr	Lys	Glu	Thr
Pro 705	Ser	Leu	Gly	Ser	Gly 710	Tyr	Asp	Pro	Tyr	Tyr 715	Thr	Tyr	Ser	Gly	Ser 720
Ile	Pro	Tyr	Leu	Asp 725	Gly	Thr	Phe	Tyr	Leu 730	Asn	His	Thr	Phe	Lys 735	Lys
Val	Ala	Ile	Thr 740	Phe	Asp	Ser	Ser	Val 745	Ser	Trp	Pro	Gly	Asn 750	Asp	Arg
Leu	Leu	Thr 755	Pro	Asn	Glu	Phe	Glu 760		Lys	Arg	Ser	Val 765	Asp	Gly	Glu
Gly	Tyr 770	Asn	Val	Ala	Gln	Cys 775	Asn	Met	Thr	Lys	Asp 780	Trp	Phe	Leu	Val
Gln 785	Met	Leu	Ala	Asn	Tyr 790	Asn	Ile	Gly	Tyr	Gln 795	Gly	Phe	Tyr	Tle	Pro 800
Glu	Ser	Tyŗ	Lys	Asp 805	Arg	Met.	Tyr	Ser	Phe 810	Phe	Arg	Asn	Phe	Gln 815	Pro
Met	Ser	Arg	Gln 820	Val	Val	Asp	Asp	Thr 825	Lys	Tyr	Lys	Glu	Tyr 830	Gln	Gln
Val	Gly		Leu				Asn 840					Val 845		Tyr	Leu
Ala	Pro 850		Met	Arg	Glụ	Gly 855	Gln	Ala	Tyr	Pro	Ala 860		Val	Pro	Tyr
Pro 865		Ile	Gly	Lys	Thr 870		Val	Asp	Ser	11e 875		Gln	Lys	Lys	Phe 880
Leu	Cys	Asp	Arg	Thr 885		Trp	Arg	Ile	Pro 890		Ser	Ser	Asn	Phe 895	Met
Ser	Met	Gly	Ala 900		Thr	Asp	Leu	Gly 905		Asn	Leu	Leu	Tyr 910		Asn

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Ser Al	a Hi:	s Al 5	a Le	u Asj	o Met	920	r Ph	e Gl	.u Va	al As	p P: 9:	ro Me 25	et A	.sp G	lu		
Pro Th	r Le	u Le	u Ty	r Va	l Le	u Pho	e Gl	u Va	al Ph	ne As 94	sp V 10	al V	al A	rg V	/al		
His GI 945 Phe Se				95	U			u Ti	nr Va 9	al Ty 55	yr L	eu A	rg T	hr I	Pro 960		
(2) II	NFORM	ITA	ON FO	R SE	Q ID	NO:	3:										
	(i) S	(A) (B) (C)	LENC TYPE STRA	CHAR STH: E: nu ANDEI OLOGY	2858 iclei NESS	bas c ac c do	e pa id ouble	airs									
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Thr"	ix)	(A)	NAM	E/KE ATIO ER I		_ 1	0 5 7		ce="}	⟨aa ⟨	can 1	pe e	ithe	er Gl	.n, H	is, o	r
	(xi)	SEQU	JENCE	DES	CRIP	TION	: SE	Q II	ON O	:3:							
ATG (_			m	mc	CG (יאכ י	ጥርር '	тст '	TAC	ATG Met	CAC His	ATC Ile 15	TCG Ser		48
GGC (CAG (SAC (GCC TAla S	rcg G Ser G	AG T	AC C	TG A	AGC Ser 25	CCC Pro	GGG Gly	CTG Leu	GTG Val	CAG Gln 30	TTT Phe	GCC Ala		96
CGC Arg	GCC A	ACC Thr 35	GAG A	ACG T	TAC I	TC F	AGC (Ser :	CTG Leu	AAT Asn	AAC Asn	AAG Lys	TTT Phe 45	AGA Arg	AAC Asn	CCC Pro		144
ACG Thr	GTG (Val.	GCG Ala	CCT Pro	ACG (CAC (GAC (Asp '	GTG Val	ACC Thr	ACA Thr	GAC Asp	CGG Arg 60	TCC Ser	CAG	CGT Arg	TTG Leu		192
ACG Thr 65	CTG Leu	CGG Arg	TTC Phe	ATC Ile	CCT Pro 70	GTG Val	GAC Asp	CGT Arg	GAG Glu	GAT Asp 75	ACT Thr	GCG Ala	TAC	TC(TAC Tyr 80	: :	_ 240
AAG Lys	GCG Ala	CGG Arg	TTC Phe	ACC Thr 85	CTA Leu	GCT Ala	GTG Val	GGT Gly	GAT Asp 90	* 1011	CGT	GTG Val	CT(G GAG 1 Asj 9	C ATO p Met 5	3	288
GCT Ala	TCC Ser	ACG Thr	TAC Tyr 100	Phe	GAC Asp	ATC Ile	CGC Arg	GGC Gly 105	val	CTG Leu	GAC Asp	AGO Aro	G GG G Gl 11	4	T AC'	r r	336
TTT Phe	AAG Lys	CCC Pro	Tyr	TCT Ser	GGC Gly	ACT Thr	GCC Ala 120	TĀT	AAC Asi	C GCC	CTO	G GC' u Ala 12		C AA	G GG 's Gl	T Y	384
GCC Ala	CCA Pro 130	AAT Asn		TGC Cys	GAA Glu	TGG Trp 135	Asp	GA/	A GC'	r GCT a Ala	r AC a Th 14		T CT a Le	T GA	AA AT Lu Il	'A .e	43

									•								
				Glu		GAT Asp						_		_	_		480
						CAC His							Tyr				528
						GGT Gly											576
						ACA Thr											624
						GAA Glu 215											672
						AAA Lys											720
						CAA Gln											768
						GAA Glu											816
	Ala		Aşn			AAC Asn										· .	864
	•					ACC Thr 295	Pro										912
		-		•		TCA Ser					•						960
						ATT Ile											1008
						GGT Gly				•				Gln			1056
					_	GTA Val			_				Thr	_			1104
						GAT Asp 375		_				Thr					1152
	Met					Val					Pro				ATT Ile 400		1200

68	
ATT GAA AAT CAT GGA ACT GAA GAT GAA CTT CCA AAT TAC TGC TTT CCA Ile Glu Asn His Gly Thr Glu Asp Glu Leu Pro Asn Tyr Cys Phe Pro 405	1248
CTG GGA GGT GTG ATT AAT ACA GAG ACT CTT ACC AAG GTA AAA CCT AAA Leu Gly Gly Val Ile Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys 420 425	1296
ACA GGT CAG GAA AAT GGA TGG GAA AAA GAT GCT ACA GAA TTT TCA GAT Thr Gly Gln Glu Asn Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp 435	1344
AAA AAT GAA ATA AGA GTT GGA AAT AAT TTT GCC ATG GAA ATC AAT CTA Lys Asn Glu Ile Arg Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu 450 455	1392
AAT GCC AAC CTG TGG AGA AAT TTC CTG TAC TCC AAC ATA GCG CTG TAT Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr 480	1440
TTG CCC GAC AAG CTA AAG TAC AGT CCT TCC AAC GTA AAA ATT TCT GAT Leu Pro Asp Lys Leu Lys Tyr Ser Pro Ser Asn Val Lys Ile Ser Asp 490	1488
AAC CCA AAC ACC TAC GAC TAC ATG AAC AAG CGA GTG GTG GCT CCC GGG Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 500	1536
TTA GTG GAC TGC TAC ATT AAC CTT GGA GCA CGC TGG TCC CTT GAC TAT Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 525	1584
ATG GAC AAC GTC AAC CCA TTT AAC CAC CAC CGC AAT GCT GGC CTG CGC Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg	1632
TAC CGC TCA ATG TTG CTG GGC AAT GGT CGC TAT GTG CCC TTC CAC ATC Tyr Arg Ser Met Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 550	1680
CAG GTG CCT CAG AAG TTC TTT GCC ATT AAA AAC CTC CTT CTC CTG CCG Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro 575	1728
GGC TCA TAC ACC TAC GAG TGG AAC TTC AGG AAG GAT GTT AAC ATG GTT Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val	<u>1</u> 776
CTG CAG AGC TCC CTA GGA AAT GAC CTA AGG GTT GAC GGA GCC AGC ATT Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile 595	1824
AAG TTT GAT AGC ATT TGC CTT TAC GCC ACC TTC TTC CCC ATG GCC CAC Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 610 620	1872
AAC ACC GCC TCC ACG CTT GAG GCC ATG CTT AGA AAC GAC ACC AAC GAC Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 635	1920
CAG TCC TTT AAC GAC TAT CTC TCC GCC GCC AAC ATG CTC TAC CCT ATA Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile 645	1968

•							•					
				CCC Pro							. •	2016
				TTC Phe 680								2064
				GAC Asp					_			2112
				TTT Phe	•							2160
				TCT Ser								2208
				GAA Glu								2256
				AAC Asn 760						GTA Val		2304
				ATT Ile						CCA, Pro		2352
				TAC Tyr						CCC Pro 800		2400
				GAT Asp					_	_		2448
				AAC Asn								2496
				CAG Gln 840								2544
				GTT Val								2592
				CGC Arg								2640
				CTG Leu			Asn			AAC Asn	·	2688
						Glu			Asp	GAG Glu	. ·	2736

70	
•	2784
CCC ACC CTT CTT TAT GTT TTG TTT GAA GTC TTT GAC GTG GTC CGT GTG Pro Thr Leu Leu Tyr Val Leu Phe Glu Val Phe Asp Val Val Arg Val 920 925	
CAC CGG CCG CAC CGC GGC GTC ATC GAA ACC GTG TAC CTG CGC ACG CCC His Arg Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro 930 935	2832
	2858
(2) INFORMATION FOR SEQ ID NO:4:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 952 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: protein	
<pre>(ix) FEATURE (A) NAME/KEY: misc_feature (B) LOCATION: 951,952 (D) OTHER INFORMATION: /note= "Xaa can be either Gln, His,</pre>	or
Thr"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser 1 5 10 15	
Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala 20 25 30	
Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro 35 40 45	
Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu 50 55 60	
Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr 80 65	
Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met 90 95	•
Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr 100 105 110	
Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly 115 120 125	
Ala Pro Asn Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile 130 135 140	
Asn Leu Glu Glu Asp Asp Asp Asn Glu Asp Glu Val Asp Glu Gln 145 150 155 160	
Ala Glu Gln Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly 165 170 175	
Ile Asn Ile Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr 180 185 190	

		•														
	Pro	Lys	Tyr 195	Ala	Asp	Lys	Thr	Phe 200	Gln	Pro	Glu	Pro	Gln 205	Ile	Gly	Glu
	Ser	Gln 210	Trp	Tyr	Glu	Thr	Glu 215	Ile	Asn	His	Ala	Ala 220	Gly	Arg	Val	Leu
٠.	Lys 225	Lys	Thr	Thr	Pro.	Met 230	Lys	Pro	Cys	Tyr	Gly 235	Ser	Tyr	Ala	Lys	Pro 240
	Thr	Asn	Glu	Asn	Gly 245	Gly	Gln	Gly	Ile	Leu 250	Val	Lys	Gln	Gln	Asn 255	Gly
	Lys	Ļeu	Glu	Ser 260	Gln	Val	Glu	Met	Gln 265	Phe	Phe	Ser	Thr	Thr 270	Glu	Ala
	Thr	Ala	Gly 275	Asn	Gly	Asp	Asn	Leu 280	Thr	Pro	Lys	Val	Val 285	Leu	Tyr	Ser
	Glu	Asp 290	Val	Asp	Ile	Glu	Thr 295	Pro	Asp	Thr	His	Ile 300	Ser	Tyr	Met	Pro
	Thr 305		Lys	Glu	Gly	Asn 310	Ser	Arg	Glu	Leu	Met 315	Gly	Gln	Gln	Ser	Met 320
	Pro	Asn	Arg	Pro	Asn 325	Tyr	Ile	Ala	Phe	Arg. 330	Asp	Asn	Phe	Ile	Gly 335	Leu
	Met	Tyr	Tyr	Asn 340	Ser	Thr	Gly	Asn	Met 345	Gly	Val	Leu	Ala	Gly 350	Gln	Ala
	Ser	Gln	Leu 355	Asn	Ala	Val	Val	Asp 360	Leu	Gln	Asp	Arg	Asn 365	Thr	Glu	Leu
	Ser	Tyr 370	Gln	Leu	Leu	Leu	Asp 375	Ser	Ile	Gly	Asp	Arg 380	Thr	Arg	Tyr	Phe
	Ser 385	Met	Trp	Asn	Gln	Ala 390	Val,	Asp	Ser	Tyr	Asp 395		Asp	Val	Arg	Ile 400
	Ile	Glu	Asn	His	Gly 405	Thr	Glu	Asp	Glu	Leu 410	Pro	Asn	Tyr	Cys	Phe 415	
	Leu	Gly	Gly	Val 420	Ile	Asn	Thr	Glu	Thr 425	Leu	Thr	Lys	Val	Lys 430	Pro	Lys
	Thr	Gly	Gln 435	Glu	Asn	Gly	Trp	Glu 440	Lys	Asp	Ala	Thr	Glu 445	Phe	Ser	Asp
	Lys	Asn 450	.Glu	Ile	Arg	Val	Gly 455	Asn	Asn	Phe	Ala	Met 460	Glu	Ile	Asn	Leu
	Asn 465	Ala	Asn	Leu.	Trp	Arg 470	Asn	Phe	Leu	Tyr	Ser 475	Asn	Ile	Ala	Leu	Tyr 480
	Leu	Pro	Asp	Lys	Leu 485	Lys	Tyr	Ser	Pro	Ser 490	Asn	Val	Lys	Ile	Ser 495	Asp
	Asn	Pro	Asn	Thr 500	Tyr	Asp	Tyr	Met	Asn 505	_	Arg	Val	Val	Ala 510	Pro	Gly
	Leu	Val ·	Asp 515	_	Tyr	Ile	Asn	Leu 520	_	Ala	Arg	Trp	Ser 525	Leu	Asp	Tyr

Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile . Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 615 ' Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile Pro Ala Asn Ala Thr Asn Val Pro Ile Ser Ile Pro Ser Arg Asn Trp Ala Ala Phe Arg Gly Trp Ala Phe Thr Arg Leu Lys Thr Lys Glu Thr Pro Ser Leu Gly Ser Gly Tyr Asp Pro Tyr Tyr Thr Tyr Ser Gly Ser Ile Pro Tyr Leu Asp Gly Thr Phe Tyr Leu Asn His Thr Phe Lys Lys Val Ala Ile Thr Phe Asp Ser Ser Val Ser Trp Pro Gly Asn Asp Arg Leu Leu Thr Pro Asn Glu Phe Glu Ile Lys Arg Ser Val Asp Gly Glu Gly Tyr Asn Val Ala Gln Cys Asn Met Thr Lys Asp Trp Phe Leu Val Gln Met Leu Ala Asn Tyr Asn Ile Gly Tyr Gln Gly Phe Tyr Ile Pro Glu Ser Tyr Lys Asp Arg Met Tyr Ser Phe Phe Arg Asn Phe Gln Pro Met Ser Arg Gln Val Val Asp Asp Thr Lys Tyr Lys Asp Tyr Gln Gln Val Gly Ile Leu His Gln His Asn Asn Ser Gly Phe Val Gly Tyr Leu Ala Pro Thr Met Arg Glu Gly Gln Ala Tyr Pro Ala Asn Phe Pro Tyr Pro Leu Ile Gly Lys Thr Ala Val Asp Ser Ile Thr Gln Lys Lys Phe

Leu 865	Cys	Asp	Arg	Thr	Leu 870	Trp	Arg	Ile	Pro	Phe 875	Ser	Ser	Asn	Phe	Met 880		
Ser	Met	Gly	Ala	Leu 885	Thr	Asp	Leu	Gly	Gln 890	Asn	Leu	Leu	Tyr	Ala 895	Asn		
Ser	Ala	His	Ala 900	Leu	Asp	Met	Thr	Phe 905	Glu	Val	Asp	Pro	Met 910	Asp	Glu		
Pro	Thr	Leu 915	Leu	Tyr.	Val	Leu	Phe 920		Val	Phe	Asp	Val 925	Val	Arg	Val		
	Arg 930	Pro	His	Arg	Gly	Val 935	Ile	Glu	Thr	Val	Tyr 940	Leu	Arg	Thr	Pro		
Phe 945	Ser	Ala	Gly	Asn	Ala 950	Xaa	Xaa						-				
(2)	INFO	RMA	MOI	FOR	SEQ	ID N	10:5:						•	٠٠.			
	(i)	•	_			CTERI			3		•					•	
		(F	3) T	PE:	nucl	leic	acio	3	-		•				•		
		•	•			ESS: line		ore									
	(ii)	MOI	LECUI	LE TY	PE:	DNA	(ger	nomi	c)								
	(xi)	SE	QUEN	CE DI	ESCRI	PTIC	ON: S	SEQ I	ID NO	0:5:		•					
										GGC Gly							48
										GAG Glu							96
										CAT His							144
										GGG Gly							192
										TAC Tyr 75							240
										TGG Trp							288
									Lys					Lys	CCA Pro		336
			Ser					Thr					Gly		TCC		384

GTT Val	Leu	GTT Val	CCG Pro	GAT Asp	GAA Glu	AAA Lys 135	GGG Gly	GTG Val	CC'	T CT o Le	T CC u Pro 14		G GT' s Va	r GA l As	C TI p Le	rG eu	432
Gln	TTC Phe	TTC Phe	TCA Ser	AAT Asn	ACT Thr 150	n CC	TCT Ser	TTC Lev	AA A As	C GA n As 15	P	G CA g Gl	A GG n Gl	C AA y As	T GC sn Al	CT La 60	480
145 ACT Thr	AAA Lys	CCA Pro	AAA Lys	Val	GTT Val	TTG Leu	TAC Tyr	AG:	r GA r Gl	.u 110	T GT	A AF	AT AT	G GA t G]	AA A Lu T 175	CC hr	528
CCA Pro	GAC Asp	ACA Thr	His	Leu		TAC Tyr	AAA Lys	CC's Pro	T G(a A	AA GO ys Gl	T GI	AT GA sp Gl	A A u As	AT T sn S	CT er	576
AAA Lys	GCT Ala	ATO Met	Let	GGI	CAA Glr	CAA Gln	TCI Sei 200	AT Me	G								603
(2)	INF			1 FOI	R SEC) ID	NO:	б :									
(2)) 51	COUE	NCE (CHARA	ACTE	RIST	ICS:									
	·		(A) : (B) :	LENG'. LYPE	TH: 2	201 a ino a	amin	o ac	ids								
			•			: lin											
						: pe			^ TF	NO:	6:						
						RIPT						ara A	Ala V	al A	Ala	Glu	
	1				5					10							
As	p Gl	u Gl	ù Gl	u G1	u As	p Gl	u As	sp G	lu (25	Glu (Glu (Glu (Glu G	30	Glu	Gln	
As	n Al				ln Al	la Th	r Ly	ys L 10	ys '	Thr	His '	Val	Tyr <i>1</i> 45	Ala	Gln	Ala	
	5	0				:	ָס		•				Gln :				
	sp As	sn A	la G	lu T	hr G	ln Al 70	la L	ys I	Pro	Val	Tyr 75	Ala	Asp	Pro	Ser	Tyr 80	
G.																	
	ln Pi	ro G	lu P	ro G	ln I 85	le G	ly G	lu S	Ser	Gln 90	Trp	Asn	Glu	Ala	Asp 95	Ala	
	sn A	la A	la G 1	ly G 00	ly A	rg V	al L	eu I	Lys 105	Lys	Thr	Thr	Pro	Met 110	Lys	Pro	
	sn A	la A yr G	la G 1	ly G 00	ly A	rg V	al L	eu I	Lys 105	Lys	Thr	Thr	Pro	Met 110	Lys		
С	sn A ys T	la A yr G	la G 1 31y S .15	ly G 00 Ser T	BS ly A	rg V la A	al L rg F	eu i Pro 120	Lys 105 Thr	Lys	Thr Pro	Thr Phe	Pro Gly 125 Lys	Met 110 Gly	Lys	Pro	
C V	sn A ys T al L ln F	yr G eu V 30	la G ly S .15	ly G 00 Ser T Pro F	ly A lyr A Asp C	rg V la A Slu I Thr T	al L rg F ys (eu : Pro 120 Sly Ser	Lys 105 Thr Val Leu	Lys Asn Pro	Thr Pro Leu Asp 155	Thr Phe Pro 140 Arg	Pro Gly 125 Lys	Met 110 Gly Val	Lys Gln Asr	Pro	

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Pro Asp Thr His Leu Ser Tyr Lys Pro Gly Lys Gly Asp Glu Asn Ser 180 185 190 Lys Ala Met Leu Gly Gln Gln Ser Met 195 (2) INFORMATION FOR SEQ ID NO:7: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 567 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7: CCT TGC GAA TGG GAT GAA GCT GCT ACT GCT CTT GAA ATA AAC CTA GAA 48 Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu 96 GAA GAG GAC GAT GAC AAC GAA GAC GAA GTA GAC GAG CAA GCT GAG CAG Glu Glu Asp Asp Asp Asn Glu Asp Glu Val Asp Glu Gln Ala Glu Gln ·25 CAA AAA ACT CAC GTA TTT GGG CAG GCG CCT TAT TCT GGT ATA AAT ATT 144 Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile . 35 40 192 ACA AAG GAG GGT ATT CAA ATA GGT GTC GAA GGT CAA ACA CCT AAA TAT Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr Pro Lys Tyr · 50 55 240 GCC GAT AAA ACA TTT CAA CCT GAA CCT CAA ATA GGA GAA TCT CAG TGG Ala Asp Lys Thr Phe Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp 65 70 75 TAC GAA ACT GAA ATT AAT CAT GCA GCT GGG AGA GTC CTT AAA AAG ACT 288 Tyr Glu Thr Glu Ile Asn His Ala Ala Gly Arg Val Leu Lys Lys Thr 85 336 ACC CCA ATG AAA CCA TGT TAC GGT TCA TAT GCA AAA CCC ACA AAT GAA Thr Pro Met Lys Pro Cys Tyr Gly Ser Tyr Ala Lys Pro Thr Asn Glu 100 105 384 AAT GGA GGG CAA GGC ATT CTT GTA AAG CAA CAA AAT GGA AAG CTA GAA Asn Gly Gly Gln Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu 115 120 432 AGT CAA GTG GAA ATG CAA TTT TTC TCA ACT ACT GAG GCG ACC GCA GGC Ser Gln Val Glu Met Gln Phe Phe Ser Thr Thr Glu Ala Thr Ala Gly 130 135 480 AAT GGT GAT AAC TTG ACT CCT AAA GTG GTA TTG TAC AGT GAA GAT GTA Asn Gly Asp Asn Leu Thr Pro Lys Val Val Leu Tyr Ser Glu Asp Val 145 155 · 150 528 GAT ATA GAA ACC CCA GAC ACT CAT ATT TCT TAC ATG CCC ACT ATT AAG Asp Ile Glu Thr Pro Asp Thr His Ile Ser Tyr Met Pro Thr Ile Lys 165 170 175

GAA GGT AAC TCA CGA GAA CTA ATG GGC CAA CAA TCT ATG

Glu Gly Asn Ser Arg Glu Leu Met Gly Gln Gln Ser Met 180

- (2) INFORMATION FOR SEQ ID NO:8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 189 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu
1 15

Glu Glu Asp Asp Asp Asn Glu Asp Glu Val Asp Glu Gln Ala Glu Gln 20 25 30

Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile 35 40 45

Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr Pro Lys Tyr 50 60

Ala Asp Lys Thr Phe Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp
65 70 75 80

Tyr Glu Thr Glu Ile Asn His Ala Ala Gly Arg Val Leu Lys Lys Thr 85 90 95

Thr Pro Met Lys Pro Cys Tyr Gly Ser Tyr Ala Lys Pro Thr Asn Glu 100 105 110

Asn Gly Gln Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu 115 120 125

Ser Gln Val Glu Met Gln Phe Phe Ser Thr Thr Glu Ala Thr Ala Gly 130

Asn Gly Asp Asn Leu Thr Pro Lys Val Val Leu Tyr Ser Glu Asp Val 145 150 150

Asp Ile Glu Thr Pro Asp Thr His Ile Ser Tyr Met Pro Thr Ile Lys
165 170 175

Glu Gly Asn Ser Arg Glu Leu Met Gly Gln Gln Ser Met 180

- (2) INFORMATION FOR SEQ ID NO:9:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 153 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ACC GAA GAT AGC GGC CGG GCA GTT GCC GAG GAT GAA GAA GAG GAA GAT

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Thr 1	Glu	Asp	Ser	Gly 5	Arg	Ala	Val	Ala	Glu 10	Asp	Glu	Glu	Glu	Glu 15	Asp			
										AAC Asn							96	;
										CCT Pro					ACA Thr		144	
_	ACA Thr 50					,				· .		. •			···		153	}
(2)	INFO	RMAI	CION	FOR	SEQ	ID N	10:10):									•	
	(i)	(<i>I</i>	A) LE 3) TY	NGTH	IARAC I: 51 amir OGY:	ami	no a	•	5		. ,			:				
	(ii)	MOI	LECUI	E TY	PE:	pept	ide									 		
	(xi)	SEC	QUENC	CE DE	SCRI	PTIC	on: S	SEQ [ED NO	0:10:	- }.							
Thr 1	Glu	Asp	Ser	Gly 5	Arg	Ala	Val	Ala	Glu 10	Asp	Glu	Glu	Glu	Glu 15	-			
Glu	Asp	Glu	Glu 20	Glu	Glu	Glu	Glu	Glu 25	Gln	Asn	Ala	Arg	Asp 30	Gln	Ala	·		
Thr	Lys	Lys 35	Thr	His	Val	Tyr	Ala 40	Gln	Ala	Pro	Leu	Ser 45	Gly	Glu	Thr		٠	
Ile	Thr 50	Lys																
(2)	INFO	ORMA	NOIT	FOR	SEQ	ID i	10:11	l :										
	(i)	(1	A) LE B) TY C) ST	ENGTI (PE: [RANI	HARAC nucl DEDNI DGY:	85 ba leić ESS:	ase p acid doub	pairs d	S									
	(ii)	MOI	LECUI	LE T	PE:	DNA	(ger	nomi	c)									
	(xi)	SE	QUENC	CE DE	ESCR	PTIC	ON: S	SEQ :	ID N	0:11								
•										GAA Glu							48	}
						•				CAA Gln						•	96	5
			Pro					Asn		ACA Thr							135	5

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu Glu Glu Asp Asp Asn 15

Glu Asp Glu Val Asp Glu Gln Ala Glu Gln Gln Lys Thr His Val Phe
20 25 30

Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile Thr Lys Glu
35 40 45

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 33 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TCA GAC AAT GCA GAA ACA CAA GCT AAA CCT GTA Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val 1 5

- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 11 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val

- (2) INFORMATION FOR SEQ ID NO:15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GTC GAA GGT CAA ACA CCT AAA Val Glu Gly Gln Thr Pro Lys 1 21

- (2) INFORMATION FOR SEQ ID NO:16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Val Glu Gly Gln Thr Pro Lys

- (2) INFORMATION FOR SEQ ID NO:17:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

AAC GAA GCT GAT GCT AAT GCG GCA Asn Glu Ala Asp Ala Asn Ala Ala 1 5 24

- (2) INFORMATION FOR SEQ ID NO:18:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Asn Glu Ala Asp Ala Asn Ala Ala 1

- (2) INFORMATION FOR SEQ ID NO:19:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

TAC GAA ACT GAA ATT AAT CAT GCA Tyr Glu Thr Glu Ile Asn His Ala

- (2) INFORMATION FOR SEQ ID NO:20:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Tyr Glu Thr Glu Ile Asn His Ala

- (2) INFORMATION FOR SEQ ID NO:21:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 42 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TCC GTT CTG GTT CCG GAT GAA AAA GGG GTG CCT CTT CCA AAG Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys 1 42

- (2) INFORMATION FOR SEQ ID NO:22:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys
1 5 10

- (2) INFORMATION FOR SEQ ID NO:23:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 42 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GGC ATT CTT GTA AAG CAA CAA AAT GGA AAG CTA GAA AGT CAA Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu Ser Gln 1

- (2) INFORMATION FOR SEQ ID NO:24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids

	01	
	(B) TYPE: amino acid (D) TOPOLOGY: linear	
-	(ii) MOLECULE TYPE: peptide	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
Gly 1	Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu Ser Gln 5 10	
(2)	INFORMATION FOR SEQ ID NO:25:	
	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 51 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
	AAT ACT ACC TCT TTG AAC GAC CGG CAA GGC AAT GCT ACT AAA CCA Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro 5 10 15	48
AAA Lys		51
(2)	INFORMATION FOR SEQ ID NO:26:	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 17 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
Ser 1	Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro 5 10 15	
Lys		
(2)	INFORMATION FOR SEQ ID NO:27:	
	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 48 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: DNA (genomic)	
•	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:	
	ACT ACT GAG GCG ACC GCA GGC AAT GGT GAT AAC TTG ACT CCT AAA Thr Thr Glu Ala Thr Ala Gly Asn Gly Asp Asn Leu Thr Pro Lys 5 10 15	4 8
(2)	INFORMATION FOR SEQ ID NO:28:	
	(i) SEQUENCE CHARACTERISTICS:	

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Ser Thr Thr Glu Ala Thr Ala Gly Asn Gly Asp Asn Leu Thr Pro Lys
1 10 15

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TTG TAC AGT GAA GAT GTA AAT ATG Leu Tyr Ser Glu Asp Val Asn Met 1

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Leu Tyr Ser Glu Asp Val Asn Met

- (2) INFORMATION FOR SEQ ID NO:31:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid(C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

TTG TAC AGT GAA GAT GTA GAT ATA Leu Tyr Ser Glu Asp Val Asp Ile 1

(2) INFORMATION FOR SEQ ID NO: 32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

24

- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Leu Tyr Ser Glu Asp Val Asp Ile
1

- (2) INFORMATION FOR SEQ ID NO:33:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GGA AAA GGT GAT GAA AAT TCT AAA GCT ATG TTG GGT Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly 1

36

- (2) INFORMATION FOR SEQ ID NO:34:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid.
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
- Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly
 1 1 10
- (2) INFORMATION FOR SEQ ID NO:35:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ACT ATT AAG GAA GGT AAC TCA CGA GAA CTA ATG GGC Thr Ile Lys Glu Gly Asn Ser Arg Glu Leu Met Gly 1

- (2) INFORMATION FOR SEQ ID NO:36:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide.
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

165

Thr Ile Lys Glu Gly Asn Ser Arg Glu Leu Met Gly
1 10

- (2) INFORMATION FOR SEQ ID NO:37:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 165 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

AAT TAT TGT TTT CCT CTT GGG GGT ATT GGG GTA ACT GAC ACC TAT CAA

Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln

1 15

GCT ATT AAG GCT AAT GGC AAT GGC TCA GGC GAT AAT GGA GAT ACT ACA Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr

TGG ACA AAA GAT GAA ACT TTT GCA ACA CGT AAT GAA ATA GGA GTG GGT
Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly
35

AAC AAC TTT GCC ATG GAA ATT Asn Asn Phe Ala Met Glu Ile 50 55

- (2) INFORMATION FOR SEQ ID NO:38:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 55 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln
1 10 15

Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr 20 25 30

Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly 35

Asn Asn Phe Ala Met Glu Ile 50 55

- (2) INFORMATION FOR SEQ ID NO:39:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 153 base pairs(B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)

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	(xi)	SEÇ	QUENC	CE DE	ESCR	PTI	ON: S	SEQ I	ID NO	0:39	:						
							GGT Gly										48
							CAG Gln										96
							GAA Glu 40										144
•	GAA Glu 50		·														153
(2)	INFO	RMAI	NOI	FOR	SEQ	ID I	NO:40):					,				
, ,	(i)	(<i>I</i>	A) LH B) TY		d: 51	l am:			.				• • •		•		
	(ii)	MOI	LECUI	LE TY	PE:	pep	tide										
	(xi)	SEÇ	QUENC	CE DE	ESCRI	PTI	: NC	SEQ I	ID NO	D:40	:		•				
Asn 1	Tyr	Cys	Phe	Pro 5	Leu	Gly	Gly	Val	Ile 10	Asn	Thr	Glu	Thr	Leu 15	Thr		
Lys	Val	Lys	Pro 20	Lys	Thr	Gly	Gln	Glu 25	Asn	Gly	Trp	Glu :	Lys 30	Asp	Ala		
Thr	Glu	Phe 35	Ser	Asp	Lys	Asn	Glu 40	Ile	Arg	Val	Gly	Asn 45	Asn	Phe	Ala	•	
Met	Glu 50	Ile	·		٠			•	· .·								
(2)	INFO	RMA	NOI	FOR	SEQ	ID !	NO: 4	1:									
	(i)	(1 (1	A) L1 B) T'(C) S'(ENGT:	H: 54 nuci DEDNI	4 ba leic ESS:	ISTIC se pa acic doul ear	airs d									-
	,(ii)	. MOI	LECU:	LE T	YPE:	DNA	(ġe	nomi	c)	•							
	(xi)	SE	QUEN	CE DI	ESCR:	IPTI	ON:	SEQ	ID N	0:41	:						
						•	ATT Ile			Asn					GGC Gly		48
	AAT Asn																54
(2)	INFO	ORMA'	TION	FOR	SEQ	ID	NO:4	2:								•	

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 amino acids

- (B) TYPE: amino acid(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly 1 5

Asp Asn

- (2) INFORMATION FOR SEQ ID NO:43:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 87 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

AAT ACA GAG ACT CTT ACC AAG GTA AAA CCT AAA ACA GGT CAG GAA AAT
Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys Thr Gly Gln Glu Asn
1 5

48

GGA TGG GAA AAA GAT GCT ACA GAA TTT TCA GAT AAA AAT Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp Lys Asn 20

87

- (2) INFORMATION FOR SEQ ID NO: 44:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys Thr Gly Gln Glu Asn

Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp Lys Asn 20

- (2) INFORMATION FOR SEQ ID NO:45:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

ACT TTT GCA ACA CGT AAT GAA
Thr Phe Ala Thr Arg Asn Glu
1 5

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- (2) INFORMATION FOR SEQ ID NO:46:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids.
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

Thr Phe Ala Thr Arg Asn Glu

- (2) INFORMATION FOR SEQ ID NO:47:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

ACA GAA TTT TCA GAT AAA AAT GAA
Thr Glu Phe Ser Asp Lys Asn Glu
1

24

- (2) INFORMATION FOR SEQ ID NO:48:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

Thr Glu Phe Ser Asp Lys Asn Glu

- (2) INFORMATION FOR SEQ ID NO:49:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: other nucleic acid
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

GAC TAC AAA GAC GAC GAC GAC AAA Asp Tyr Lys Asp Asp Asp Asp Lys

- (2) INFORMATION FOR SEQ ID NO:50:
 - (i) SEQUENCE CHARACTERISTICS:

		(B)	LENC TYPE TOP	E: an	nino	acid	i	ds								
I	(ii)	MOLE	CULE	TYP	E: pe	eptio	de									
	(xi)	SEQU	ENCE	DES	CRIPT	rion	: SE	Q ID	NO:	50:						
Asp '	Tyr 1	Lys A	A qa	sp A	sp As	sp L	ys									
(2)	INFO	RMAT	ON F	OR S	EQ I	D NO	:51:			٠						
	(i)	(A) (B) (C)	JENCE LEN TYP STR TOP	GTH: E: n ANDE	290 ucle DNES	7 ba ic a S: d	se p cid loubl	airs								
	(ii)	MOL	ECULE	TYE	PE: D	NA (geno	omic)								
	(xi)	SEQ	UENCE	E DES	SCRIP	MOIT	1: SE	EQ II	NO:	51:						
ATG	GCT Ala	ACC Thr	CCT S	rcg <i>l</i> Ser h	Met M	iet i	CCG (olii .	rgg T	CT Ser S	TAC A	ATG (CAC A	ATC Ile	TCG Ser 15	48
GGC Gly	CAG Gln	GAC Asp	GCC '	TCG (Ser (כאכ יי	ኮክሮ (ጉጥር ነ	AGC (CCC (Pro (GGG Gly	CTG (Leu	GTG (Val	CAG Gln	TTT Phe 30	GCC Ala	96
CGC Arg	GCC Ala	ACC Thr	GAG Glu 35	ACG Thr	TAC T	TTC I	AGC Ser	CTG Leu 40	AAT / Asn /	AAC Asn	AAG Lys	TTT Phe	AGA Arg 45	AAC Asn	CCC Pro	144
ACG Thr	GTG Val	GCA Ala 50	CCT Pro	ACG Thr	CAC His	GAC Asp	GTA Val 55	ACC Thr	ACA Thr	GAC Asp	CGG Arg	TCC Ser 60	CAG Gln	CGT Arg	TTG Leu	192
ACG Thr	CTG Leu 65	Arg	TTC Phe	ATC Ile	CCT Pro	GTG Val 70	GAC Asp	CGC Arg	GAG Glu	GAT Asp	ACC Thr 75	GCG Ala	TAC Tyr	TCG Ser	TAC Tyr	240
AAA Lys	. Ala	CGG Arg	TTC Phe	ACC Thr	CTG Leu 85	GCT Ala	GTG Val	GGT Gly	GAC Asp	AAC Asn 90	CGT Arg	GTG Val	CTT Leu	GAT Asp	ATG Met 95	288
GCT Ala	TCC a Ser	C ACG	TAC	TTT Phe 100	Asp	ATC Ile	CGC Arg	GGC	GTG Val 105	CTG Leu	GAC Asp	AGG Arg	GGG Gly	CCT Pro	ACT Thr	336
TT'	r AA(G CCC	TAC Tyr	Ser	GGC Gly	ACT Thr	GCC Ala	TAC Tyr 120	Asn	GCT	CTA Leu	GCT Ala	Pro 125		G GGC G Gly	384
GC Al	T CC a Pr	T AA(o Asi 13(n Ser	TGT Cys	GAG Glu	TGG	GAA Glu 135	GIN	ACC Thr	GAA Glu	A GAT 1 Asp	AGC Ser 140		C CGO y Aro	g GCA g Ala	432
. GT Va	T GC	a Gl	G GAT u Asp	GAA	A GAA 1 Glu	GAG Glu 150	1 GT	A GAT 1 Asp	GAA	A GAT	r GAF o Glu 155		A GAG	G GA u Gl	A GAA u Glu	480

			CGA Arg 165										528
			TCT Ser			Thr				Gly	Gln		576
			GCA Ala									•	624
			GAA Glu										672
			GCA Ala										720
			GGA Gly 245								GGT Gly 255		768
			GTT Val										816
	•		TTC Phe									*	864
		Thr	CCA Pro									<i>:</i>	912
			ACA Thr						_				960
			ATG Met 325										1008
			AGG Arg										1056
			GGT Gly										1104
			CAA										1152
			GGT Gly		Arg	_							1200
Ala			TAT Tyr 405	Asp				Ile			CAT His 415	٠.	1248

											90										
	GGA A	ACT Thr	GAG Glu	GA' Asj	b G1	AA T Lu L 20	TG (eu F	CCA . Pro	AAT Asn	TAT Tyr	TGT Cys 425		CC Pr	T C	TT G eu G	_	GGT Gly 430	ATT Ile		1296	
	GGG (GTA Val	ACT Thr	GA As:	p Ti	CC T	AT (CAA Gln	GCT Ala	ATT Ile 440	AAG Lys	GC'S	r AA a As	T G	-y -	AAT Asn 445	GGC Gly	TCA Ser	\	1344	
	GGC (GAT Asp	AAT Asr 450	r GG n Gl	n C	AT A sp T	ACT I	ACA Thr	TGG Trp 455	ACA Thr	AAF Lys	A GA'	r GA p Gl	Lu	CT Thr 1	TTT Phe	GCA Ala	AC#	A C	1392	
	CGT Arg	AAT Asn 465	GAZ Gl:		A G	GA (Val	GGT Gly 470	AAC Asn	AAC Asn	TTT	r GC e Al	a m	rG G et G 75	SAA I	ATT Ile	AAC Asn	CT	A u	1440	
•	AAT Asn 480			C CI n Le	TA T eu T	rp l	AGA Arg 485	AAT Asn	TTC Phe	CTT Leu	TAC Ty:	C TC r Se 49	T A	AT A	ATT [le	GCG Ala	CTG Leu	TA Ty	C r 5	1488	
	CTG Leu	CCA Pro	GA As	C A	ys I	TA deu	AAA Lys	TAC Tyr	AAC Asn	CCC	AC Th	T M	AT G	TG (GAA Glu	ATA Ile	TCT Ser 510		C P	1536	
	AAC Asn	CCC	: AA > As	in T	00 M	n x 🗸	GAC Asp	TAC Tyr	ATC Met	AAC Ası 520	тъй	G CO	SA G	TG (GTG Val	GCT Ala 525		: GG : G1	G . Y	1584	
	CTT Leu	GT#	L As			TAC Tyr	ATT Ile	AAC Asn	CTI Lev 539	I GT	g GC y Al	CG Co La A	GC I	GG Trp	TCT Ser 540		GAC Asp	TA Ty	AC yr	1632	
	ATG Met	GAG Asj	C Al		TT Z	AAT Asn	CCC Pro	TTI Phe 550	T AAG ASI	C CA n Hi	C CA s Hi	AC C is A	ry r	AAT Asn 555	GCG Ala	GGC Gly	CTO	C CC	GT rg	1680	
	Tyr	CG Ar		CC A	ATG Met	TTG Leu	TTG Leu 565	L GT	A AA y As	C GG n Gl	C CC	rg i	AC (yr)	GTG Val	CCC Pro	TTI Phe	CA Hi	C A s I 5	TT le 75	1728	
	560 CAG Gln		G C 1 P	CC (CAA Gln	AAG Lys 580	TTI Phe	, արար	T GC e Al	C AT a Il	.е ь	AA A ys A	AC	CTC Leu	CTC	CTO	C CT 1 Le 59	G C u P 0	CA ro	1776	
	GGC Gl	TC Y Se	A Ter T	yr '	ACA Thr 595	~ ~ ~ ~	C 7. 7	A TG	G AA	in Pi	rc A ne A	GG A	AAG Lys	GAT Asp	GTI Val	AA L As:		'G G et V	TT Val	1824	
	CT(Let	G CF	ln S			CTG Leu	G GGI	A AA y As	C GF	AT C' sp L	rr A eu A	AGA (GTT Val	GAC Asp	GG(Gl ₂ 62)	<i>y</i>	T AG a Se	GC A	TT [le	1872	
	AA(Ly:	s Pl	TT (AGC Ser	ATT	TG Cy	s Le	T T	NC G	CC <i>l</i>	ACC Thr	TTC Phe	TTC Phe 635		C AT	G G(CC (CAC His	1920	
	As	C A	25 CG (hr)	GCC Ala	TCC Ser	ACC Th:	G CT r Le 64	G GI	AA G lu A	CC A la M	TG (CTC Leu	AGA Arg 650	V	r GA n As	C AC	CC A		GAC Asp 655	1968	
	64		CC	TTT	AAT	GA	с пл	יר כי	TT T	CC C	SCC	GCC Ala	AAC	AT(G CI t Le	ra Tr	AC C	CC	ATA Ile	2016	

CCC GCC AAC GCC ACC AAC GTG CCC ATC TCC ATC CCA TCG CGC AAC TGG 2064 Pro Ala Asn Ala Thr Asn Val Pro Ile Ser Ile Pro Ser Arg Asn Trp	
675 680 685	
GCA GCA TTT CGC GGT TGG GCC TTC ACA CGC TTG AAG ACA AAG GAA ACC 2112 Ala Ala Phe Arg Gly Trp Ala Phe Thr Arg Leu Lys Thr Lys Glu Thr 690 695 700	·
CCT TCC CTG GGA TCA GGC TAC GAC CCT TAC TAC ACC TAC TCT GGC TCC 2160 Pro Ser Leu Gly Ser Gly Tyr Asp Pro Tyr Tyr Thr Tyr Ser Gly Ser 705 710 715	
ATA CCA TAC CTT GAC GGA ACC TTC TAT CTT AAT CAC ACC TTT AAG AAG 2208 Ile Pro Tyr Leu Asp Gly Thr Phe Tyr Leu Asn His Thr Phe Lys Lys 720 725 730 735	•
GTG GCC ATT ACC TTT GAC TCT TCT GTT AGC TGG CCG GGC AAC GAC CGC 2256 Val Ala Ile Thr Phe Asp Ser Ser Val Ser Trp Pro Gly Asn Asp Arg 740 745 750	·
CTG CTT ACT CCC AAT GAG TTT GAG ATT AAA CGC TCA GTT GAC GGG GAG 2304 Leu Leu Thr Pro Asn Glu Phe Glu Ile Lys Arg Ser Val Asp Gly Glu 755 760 765	
GGC TAC AAC GTA GCT CAG TGC AAC ATG ACC AAG GAC TGG TTC CTG GTG 2352 Gly Tyr Asn Val Ala Gln Cys Asn Met Thr Lys Asp Trp Phe Leu Val 770 775 780	
CAG ATG TTG GCC AAC TAC AAT ATT GGC TAC CAG GGC TTC TAC ATT CCA 2400 Gln Met Leu Ala Asn Tyr Asn Ile Gly Tyr Gln Gly Phe Tyr Ile Pro 785 790 795	
GAA AGC TAC AAG GAC CGC ATG TAC TCG TTC TTC AGA AAC TTC CAG CCC 2448 Glu Ser Tyr Lys Asp Arg Met Tyr Ser Phe Phe Arg Asn Phe Gln Pro 800 805 810 815	
ATG AGC CGG CAA GTG GTT GAC GAT ACT AAA TAC AAG GAG TAT CAG CAG 2496 Met Ser Arg Gln Val Val Asp Asp Thr Lys Tyr Lys Glu Tyr Gln Gln 820 825 830	
GTT GGA ATT CTT CAC CAG CAT AAC AAC TCA GGA TTC GTA GGC TAC CTC 2544 Val Gly Ile Leu His Gln His Asn Asn Ser Gly Phe Val Gly Tyr Leu 835 840 845	·
GCT CCC ACC ATG CGC GAG GGA CAG GCT TAC CCC GCC AAC GTG CCC TAC 2592 Ala Pro Thr Met Arg Glu Gly Gln Ala Tyr Pro Ala Asn Val Pro Tyr 850 855 860	
CCA CTA ATA GGC AAA ACC GCG GTT GAC AGT ATT ACC CAG AAA AAG TTT 2640 Pro Leu Ile Gly Lys Thr Ala Val Asp Ser Ile Thr Gln Lys Lys Phe 865 870 875	
CTT TGC GAT CGC ACC CTT TGG CGC ATC CCA TTC TCC AGT AAC TTT ATG 2688 Leu Cys Asp Arg Thr Leu Trp Arg Ile Pro Phe Ser Ser Asn Phe Met 880 885 890 895	
TCC ATG GGC GCA CTC ACA GAC CTG GGC CAA AAC CTT CTC TAC GCC AAC 2736 Ser Met Gly Ala Leu Thr Asp Leu Gly Gln Asn Leu Leu Tyr Ala Asn 900 905 910	
TCC GCC CAC GCG CTA GAC ATG ACT TTT GAG GTG GAT CCC ATG GAC GAG 2784 Ser Ala His Ala Leu Asp Met Thr Phe Glu Val Asp Pro Met Asp Glu 915 920 925	
	•

92	
CCC ACC CTT CTT TAT GTT TTG TTT GAA GTC TTT GAC GTG GTC CGT GTG Pro Thr Leu Leu Tyr Val Leu Phe Glu Val Phe Asp Val Val Arg Val 930 935	2832
CAC CAG CCG CAC CGC GGC GTC ATC GAG ACC GTG TAC CTG CGC ACG CCC His Gln Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro 945 955	2880
TTC TCG GCC GGC AAC GCC ACA ACA TAA Phe Ser Ala Gly Asn Ala Thr Thr 960 965	2907
(2) INFORMATION FOR SEQ ID NO:52:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 967 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: protein	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:	
Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser Gly 1 10 15	
Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala Arg 20 25 30	
Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro Thr 35 40 45	
Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu Thr 50 55 60	
Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr Lys 65 70 75 80	
Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met Ala 85 90 95	
Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr Phe 100 105 110	
Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly Ala 115 120 125	٠
Pro Asn Ser Cys Glu Trp Glu Gln Thr Glu Asp Ser Gly Arg Ala Val 130 135 140	
Ala Glu Asp Glu Glu Glu Glu Asp Glu	
Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr Ala 165 170 175	
Gln Ala Pro Leu Ser Gly Glu Thr Ile Thr Lys Ser Gly Leu Gln Ile 180 185 190	
Gly Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val Tyr Ala Asp Pro 195 200 205	
Ser Tyr Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp Asn Glu Ala 210 215 220	

Asp 225	Ala	Asn	Ala	Ala	Gly 230	Gly	Arg	Val	Leu	Lys 235	Lys	Thr	Thr	Pro	Met 240	. •		•				
Lys	Pro	Cys	Tyr	Gly 245	Ser	Tyr	Ala	Arg	Pro 250	Thr	Asn	Pro	Phe	Gly 255	Gly			•• •	· ·		٠.	
Gln	Ser	Val	Leu 260	Val	Pro	Asp	Glu	Lys 265	Gly	Val	Pro	Leu	Pro 270	Lys	Val							
Asp	Leu	Gln 275	Phe	Phe	Ser	Asn	Thr 280	Thr	Ser	Leu	Asn	Asp 285	Arg	Gln	Gly						_	
Asn	Ala 290	Thr	Lys	Pro	Lys	Val 295	Val _.	Leu [.]	Tyr	Seŗ	Glu 300	Asp	Val	Asn	Met							
Glu 305	Thr	Pro	Asp	Thr	His 310	Leu	Ser	Tyr	Lys	Pro 315	Gly	Lys.	Gly	Asp	Glu 320		•			÷		
Asn	Ser	Lys	Ala	Met 325	Leu	Gly	Gln	Gln	Ser 330	Met	Pro	Asn	Arg	Pro 335	Asn							
Tyr	Ile	Ala	Phe 340	Arg	Asp	Asn	Phe	11e 345	Gly	Leu	Met	Tyr	Tyr 350	Asn	Ser				•		•	
Thr	Gly	Asn 355	Met	Gly	Vai	Leu	Ala 360	Gly	Gln	Ala	Ser	Gln 365	Leu	Asn	Ala							•
Val	Val 370	Asp	Leu	Gln	Asp	Arg 375	Asn	Thr	Glu	Leu	Ser 380	Tyr	Gĺn	Leu	Leu.		··.	• • •				
Leu 385	Asp	Ser	Ile	Gly	Asp 390	Arg	Thr	Arg	Tyr	Phe 395	Ser	Met	Trp	Asn	Gln 400	.**	٠.					
Ala	Val	Asp	Ser	Tyr 405	Asp	Pro	Asp	Val	Arg 410	Ile	Ile	Glu	Asn	His 415	Gly							
Thr	Glu	Asp	Glu 420	Leu	Pro	Asn	Ťyr	Cys 425	Phe	Pro	Leu	Gly	Gly 430	Ile	Gly				•			
Val	Tḥr	Asp 435	Thr	Tyr	Gln	Ala	Ile 440	Lys	Ala	Asn	Gly	Asn 445	Gly	Ser	Gly							
Asp	Asn 450	Gly	Asp	Thr	Thr	Trp 455	Thr	Lys	Asp	Glu	Thr 460		Ala	Thr	Arg							
Asn 465	Glu	Ile	Gly	Val	Gly 470	Asn	Asn	Phe	Ala	Met 475	Glu	Ile	Asn	Leu	Asn 480					٠		
Ala	Asn	Leu	Trp	Arg 485	Asn	Phe	Leu	Tyr	Ser 490		Ile	Ala	Leu	Tyr 495	Leu						,	
Pro	Asp	Lys	Leu 500	Lys	Tyr	Asn	Pro	Thr 505	Asn	Va1	Glu	Ile	Ser 510	Asp	Asn							
Pro	Asn	Thr 515	Tyr	Asp	Tyr	Met	Asn 520	Lys	Arg	Val	Val	Ala 525	Pro	Gly	Leu							
Val	Asp 530	Суѕ	Tyr	Ile	Asn	Leu 535	Gly	Ala	Arg	Trp	Ser 540		Asp	Tyr	Met							
Asp 545	Asn	Val	Asn	Pro	Phe 550		His	His	Arg	Asn 555		Gly	Leu	Arg	Tyr 560			· · · .	•		•	
					- -																	
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Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile Pro Ala Asn Ala Thr Asn Val Pro Ile Ser Ile Pro Ser Arg Asn Trp Ala Ala Phe Arg Gly Trp Ala Phe Thr Arg Leu Lys Thr Lys Glu Thr Pro Ser Leu Gly Ser Gly Tyr Asp Pro Tyr Tyr Thr Tyr Ser Gly Ser Ile Pro Tyr Leu Asp Gly Thr Phe Tyr Leu Asn His Thr Phe Lys Lys Val Ala Ile Thr Phe Asp Ser Ser Val Ser Trp Pro Gly Asn Asp Arg Leu Leu Thr Pro Asn Glu Phe Glu Ile Lys Arg Ser Val Asp Gly Glu Gly Tyr Asn Val Ala Gln Cys Asn Met Thr Lys Asp Trp Phe Leu Val Gln Met Leu Ala Asn Tyr Asn Ile Gly Tyr Gln Gly Phe Tyr Ile Pro Glu Ser Tyr Lys Asp Arg Met Tyr Ser Phe Phe Arg Asn Phe Gln Pro Met Ser Arg Gln Val Val Asp Asp Thr Lys Tyr Lys Glu Tyr Gln Gln Val Gly Ile Leu His Gln His Asn Asn Ser Gly Phe Val Gly Tyr Leu Ala Pro Thr Met Arg Glu Gly Gln Ala Tyr Pro Ala Asn Val Pro Tyr Pro Leu Ile Gly Lys Thr Ala Val Asp Ser Ile Thr Gln Lys Lys Phe Leu Cys Asp Arg Thr Leu Trp Arg Ile Pro Phe Ser Ser Asn Phe Met Ser

Met	Gly	Ala	Leu 900	Thr	Asp	Leu	Gly	Gln 905	Asn	Leu	Leu	Tyr	Ala 910	Asn	Ser	, · ·	
Ala	His	Ala 915	Leu	Asp	Met	Thr	Phe 920	Glu	Val	Asp	Pro	Met 925		Glu	Pro		
Thr	Leu -930	Leu	Tyr	Val	Leu	Phe 935	Glu	Val	Phe	Asp	Val 940	Val	Arg	Val	His		
Gln 945	Pro	His	Arg	Gly	Val 950	Ile	Glu	Thr	Val	Tyr 955	Leu	Arg	Thr	Pro	Phe 960		
Ser	Ala	Gly	Asn	Ala 965	Thr	Thr			٠						•		
(2)	INFO	ORMA	MOI	FOR	SEQ	ID N	10:53	3: •									
	(i)	(<i>I</i> (E	A) LE 3) TY C) ST	ENGTI (PE: [RANI	HARAC H: 28 nucl DEDNE DGY:	858 k Leic ESS:	acio doub	pai:	cs								
	(ii)	MOI	LECUI	LE TY	PE:	DNA	(ger	nomic	:) ·		· .						•
	(xi)	SEÇ	QUENC	CE DE	ESCRI	PTIC	on: s	SEQ I	ID NO	53:	:						
ATG					ATG Met 5												18
					GAG Glu												∂ 6
					TAC Tyr											14	14
					CAC											19	92
					CCT Pro											. 24	10
					CTA Leu 85											28	38
					GAC Asp											33	36
					GGC Gly				Asn							. 38	84
					GAA Glu											4:	32

AAC CTA GAA GAA GAG GAC GAT GAC AAC GAA GAC GAA GTA GAC GAG CAA

											9	0									
As		Leu 145	Glu	Gl	u G	lu i	Asp	Asp 150	Asp	As	n G	lu i	Asp	Glu 155	ı Va	al A	Asp	Glu	Gl	n	
A.	CT la 60	GAG Glu	CAG	CA Gl	A A n L	ıys	ACT Thr 165	CAC His	GTA Val	TT Ph	T G e G	TA .	CAG Gln 170	GC0 Ala	G CC	cT t	TAT Tyr	TCT Ser	GG Gl 17	3	528
A	TA le	AAT Asn	ATT Ile	AC Th	ir I	AAG Lys 180	GAG Glu	GGT Gly	ATT	CA Gl	.n ı	TA le .85	GGT Gly	GT(C GZ	AA lu	GGT Gly	CAA Gln 190	AC Th	CA nr	576
C P	CT ro	AAA Lys	TA? Ty:	c Al	CC (La <i>1</i> 95	GAT Asp	AAA Lys	ACA Thr	TTI Phe	CF G]	Ln E	CT Pro	GAA Glu	CC'	T C	AA ln	ATA Ile 205	GGA Gly	GI GI	AA Lu	624
T	CT	CAG Gln	TG(Tr)	P T	AC (GAA Glu	ACT Thr	GAA Glu	ATT	e As	AT (sn i	CAT	GCA Ala	GC Al	a G	GG ly 20	AGA Arg	GTC Val	. Ci	rT eu	672
P I	AAA .ys	AAG Lys 225	AC'	יע ייי	CC (CCA Pro	ATG Met	AAA Lys	Pr	A TO	GT '	TAC Tyr	GGT Gly	TC Se 23	т т	AT Yr	GCA Ala	AAA Lys	C P	CC TO	720
י י	ACA Ihr 240	AAT Asr		A A u A	AT .sn	GGA Gly	GGG Gly 245	Gli	A GG	C A	TT le	CTT Leu	GTA Val 250	ר די	G C	CAA Gln	CAA Gln	AA1 Asr		GA 1y 55	. 768
		- CM1	A GA ı Gl	A A u S	GT	CAA Gln 260	GTG Val	GAI	A AT L Me	G C t G	AA 1n	TTT Phe 265	Pne	C TO	CA A	ACT Thr	ACT	GA(Gl)		CG la	816
	ACC Thr	GC:	A GO a Gl	y P	AAT Asn 275	GGT Gly	GAI Asp	AA :	C TT n Le	u T	CT Chr 280	CCT Pro	AA Ly:	A G'	TG (GTA Val	TTC Lev 285	3	C A	AGT Ser	864
	GA <i>F</i> Glu	A GA 1 As	p Va	TA (GAT Asp	ATA Ile	A GAM	A AC ı Th	C CC r Pi 29	O F	Asp	ACT Thr	CA:	T A' s I	16	TCT Ser 300	- Y -	C AT	G (CCC Pro	912
	AC: Thi	r AT r Il 30	T AZ	N.C. (GAA Glu	GGT Gly	r AA y As:	C TC n Se 31	r A	GA (GAA Glu	CTA	A AT n Me	it G	GC ly 15	CA <i>P</i> Glr	A CAI	A TC n Se	T I	ATG Met	960
	CCC Pri	C AA	<i>C</i> 7.	GG rg	CCT Pro	AA! Ası	T TA n Ty 32	r Il	T G	CT '	TTT Phe	AG(G GA g As	\mathbf{p}_{F}	AT Asn	TT? Phe	r AT	T GG e Gl	٠, ٦	CTA Leu 335	1008
		~ M7	AT T	AC yr	AAC Asn	AG Se:	C AC r Th	G G(r G)	ST A Ly A	AT sn	ATG Met	GG' G1 34	y va	TT (CTG Leu	GC(G GG a Gl	y C.	AA Ln 50	GCA Ala	1056
	TC Se	G C	AG T ln L	TG eu	AAT Asn 355	a Al	T GI a Va	T G	ra G al A	AT .sp	TTG Leu	GT	A GA	AC I	AGA Arg	AA As	C AC n Th	11 0.	AG lu	CTT Leu	1104
	TC Se	CA T	yr (CAG Gln	CTP	ጉ ጥጥ	G Ci	TT G	sp S	CC Ser	ATT Ile	GG Gl	T G	AT .	AGA Arg	AC Th	T M	GG T.	AC yr	TTT Phe	1152
	T(er M	mc r	r.c.c	AA' Asi	T CA	AG G Ln A	la Ņ	TT (GAC Asp	AG0 Set	C TA	AT G	sp	CCA Pro 395) As	AT G	IT A al A	.GA .rg	ATT Ile	1200
	A'			AAT	CA	T GO	GA A			GAT	GA	A C	rr C	CA	TAA	TA	AC T	GC I	TT	CCA	1248

										J ,							
	Ile 400	Glu	Asn	His	Gly	Thr 405	Glu	Asp	Glu	Leu	Pro 410	Asn	Tyr	Суз	Phe	Pro 415	
							ACA Thr										1296
							TGG Trp						Glu				1344
							GGA Gly										1392
							AAT Asn 470										1440
							TAC Tyr										1488
	AAC Asn	CCA Pro	AAC Asn	ACC Thr	TAC Tyr 500	GAC Asp	TAC Tyr	ATG Met	AAC Asn	AAG Lys 505	CGA Arg	GTG Val	GTG Val	GCT Ala	CCC Pro 510	GGG Gly	1536
							AAC Asn									TAT Tyr	1584
							TTT Phe										1632
							GGC Gly 550										1680
•							TTT Phe										1728
							TGG Trp										1776
							AAT Asn		Leu		Val	Asp	Gly				1824
							CTT Leu		•								1872
							GAG Glu 630										1920
							CTC Leu										1968

												98											
I	CCC Pro	GCC Ala	A!	AC (GCT Ala	ACC Thr 660	AA(Asr	GT V	rg (CCC Pro	ATA Ile	TC Se 66	I	ATC [le	CCC	T(CC (CGC Arg	AAC Asn 670		GG CP	•	2016
(GCG Ala	GCT Ala	T'	TC he	CGC Arg 675	GGC Gly	TG(G G(CC '	TTC Phe	ACG Thr 680	CG	sc (CTT Leu	AAC Lys	A T	CT hr	AAG Lys 685	GAA Glu	A T	cc nr		2064
•	CCA Pro	TCF Ser	L	TG eu 90	GGC Gly	TCG Ser	GG(C T	yr	GAC Asp 695	CCT Pro	TA	AT '	TAC Tyr	ACC Thi		AC yr 00	TCT Ser	GGC	T	CT er		2112
	ATA Ile	CCC Pro	T c	AC	CTA Leu	GAT Asp	GG Gl	y T	CC hr 10	TTT Phe	TAC	C'	rC eu	AAC Asn	CAC His	2 I	CC hr	TTT Phe	AA(Lys	S A	AG ys		2160
	GTG Val 720	GC Al	C A	TT le	ACC Thr	TTT Phe	GA As	p S	CT er	TCT Ser	GTC Val	: A	GC er	TGG Trp 730	PL	T G	GC Gly	AAT Asn	GA(C C p A	GC rg 35		2208
	CTG Leu	CT Le	T A	ACC Thr	CCC	AA(Asi 74(ı Gl	G I	TT	GAA Glu	ATI	; L	AG ys 45	CGC Arg	TC Se	A (GTT Val	GAC Asp	GG G1 75		AG Slu		2256
	GGT Gly	TA Ty	C I	AAC Asn	GTT Val 755	AL	C CF a Gl	lG T	rGT Cys	AAC Asn	ATO Met 760		CC hr	AAA Lys	GA As	C :	TGG Trp	TTC Phe 765		G (GTA Val		2304
	CAA Glr	AT Me	t	CTA Leu 770	Ala	AA 7 a As	C T	AC I	AAC Asn	ATT	GGG GGI	y j	AC Yr	CA(G GG	L X	TTC Phe 780	- 1 -	r AT	c (CCA Pro		2352
	GA0	ı Se	SC er	TAC Tyr	AA(G GA s As	C Cop A	rg	ATG Met 790	Ty.	C TC r Se	C T	TTC Phe	TT'	5 A.	GA rg 95	AAC Asn	TTO Pho	c CA e Gl	AG Ln	CCC Pro		2400
	ATO Med 800	G A	20	CGT Arg	CA G Gl	G GI n Va	l V	TG al 05	GAT Asp	GA'	T AC p Th	T i	AAA Lys	TA Ty 81	r 1	AG ys	GAC Asp	TA Ty	C CA	AA ln	CAG Gln 815		2448
			GC ly	ATO	C CT e Le	u Hi	AC C	AA ln	CAC His	AA As	C AA	in	TCI Ser 825	GI	A T y P	TT he	GT'	r GG 1 Gl	C TAY TO	AC yr 30	CTT Leu		2496
0	GC Al	C C a P	CC ro	ACC Th:	C AT r Me 83	t A	sc o	AA Slu	GG!	A CA y Gl	n A.	CC La	TAC Tyı	C CC	T G	CT	AA	C TI n Ph 84	-	CC ro	TAT Tyr		2544
	CC Pr	G C	TT	AT. 11 85	e Gl	SC A	AG A	ACC Thr	GC/ Al	A GI a Va 85	IT A	AC sp	AG(Se:	C AT	T A Le I	ACC Thr	CA Gl 86	ر ۱۰	AA A ys L	AG ys	TTT		2592
	CT Le	eu (GC Cys 365	GA As	m <i>C</i> /	GC A	CC (CTT Leu	TG Tr 87	p A	GC A	TC le	CC. Pr	A T	16 .	rcc Ser 875	50	T A	AC I	TT he	ATG Met		2,640
	Se				SC G y A	CA C la I	eu	ACA Thr 885	As	C C	TG G eu G	GC 1 y	CA Gl	n A	AC (sn)	CTT Leu	CI Le	C T	AC (yr <i>l</i>	GCC Ala	AAC Asn 895		2688
	_		GCC Ala	C CA	AC G is A	la 1	TA Leu 900	GAC Asp	TA : Me	CG A	CT I	TT Phe	GA G1 90	.u v	TG al	GAT Asp	r co p P:	CC A		GAC Asp 910	GAG Glu		2736

										99								
								TTT Phe										2784
								ATC Ile 935									:	2832
						GCA Ala		CAT His	AA									2858
,	(2)	INFO	ORMAT	цой	FOR	SEQ	ID N	NO:54	4:									
		(i)	(<i>I</i>	A) LE B) T	ENGTI (PE:		ol an			ds								
		(ii)	MO1	LECUI	LE TY	PE:	prot	cein	•									
		·(xi)	SEC	QUENC	CE DI	ESCRI	PTIC	on: s	SEQ I	D NO):54:	:						
	Ala 1	Thr	Pro	Ser	Met 5	Met	Pro	Gln	Trp	Ser 10	Tyr	Met	His	Ile	Ser 15	Gly	•	
	Gln	Asp	Ala	Ser 20	Glu	Tyr	Leu	Ser	Pro 25	_	Leu	Val	Gln	Phe 30	Ala	Arg		,
	Ala	Thr	Glu 35	Thr	Tyr	Phe	Ser	Leu 40	Asn	Asn	Lys	Phe	Arg 45	Asn	Prò	Thr		
	Val	Ala 50	Pro	Thr	His	Asp	Val 55	Thr	Thr	Asp	Arg	Ser 60	Gln	Arg	Leú	Thr		
	Leu 65	Arg	Phe	Ile	Pro	Val 70	Asp	Arg	Glu	Asp	Thr 75	Ala	Tyr	Ser	Tyr	Lys 80		
	Ala	Arg	Phe	Thr	Leu 85	Ala	Val	Gly	Asp	Asn 90	Arg	Val	Leu	Asp	Met 95	Ala		
	Ser	Thr	Tyr	Phe 100	Asp	Ile	Arg	Gly	Val 105	Leu	Asp	Arg	Gly	Pro 110	Thr	Phe		
	Lys	Pro	Tyr 115	Ser	Gly	Thr	Ala	Tyr 120	Asn	Ala	Leu	Ala	Pro 125	Lys	Gly	Ala		•
	Pro	Asn 130	Pro	Cys	Glu	Trp	Asp 135	Glu	Ala	Ala	Thr	Ala 140	Leu	Glu	Ile	Asn		
	Leu 145	Glu	Glu	Glu	Asp	Asp 150	Asp	Asn	Glu	Asp	Glu 155	Val	Asp	Glu	Gln	Ala 160		
	Glu	Gln	Gln	Lys	Thr 165	His	Val	Phe	Gly	Gln 170	Ala	Pro	Tyr	Ser	Gly 175	Ile		
	Asn	Ile	Thr	Lys 180	Glu	Gly	Ile	Gln	Ile 185	Gly	Val	Glu	Gly	Gln 190	Thr	Pro		
	Lys	Tyr	Ala 195	_	Ļys	Thr	Phe	Gln 200	Pro	Glu	Pro	Gln	Ile 205	Gly	Glu	Ser		
	Gln	Trp 210	Tyr	Glu	Thr	Glu	Ile 215	Asn	His	Ala	Ala	Gly 220	Arg	Val	Leu	Lys		

	Thr	Thr	Pro	Met	Lys	Pr	· o C	Cys	Ty:	r G	Sly	Ser 235	Tyr	A.	la	Lys	Pr	o I	hr 40	
225	6 1	N a w	Gly	Clu	230		v 1	[]e	Lei	u V			Glr	G.	ln	Asn	G]	Ly I	ıys	
				245							250									
	,		Gln 260						20	5										
Ala	Gly	Asn 275	Gly	Asp	Ası	ı Le	eu '	Thr 280	Pr	0]	Lys	Val	Va:	L L 2	eu 85	Tyr	S	er (Slu	ł
Asp	Val 290	Asp	Ile	Glu	Th:	r Pi	ro / 95	Asp	Th	r :	His	Ile	Se:	r T O	yr	Met	. P.	ro '	Thr	:
Ile 305	Lys	Glu	Gly	Asr	Se. 31	r A:	rg	Glu	Le	eu	Met	Gly 315	Gl	n G	ln	Ser	: M	et	Pro 320	0
Asn	Arg	Pro	Asn	Ту: 32	r Il	e A	la	Phe _.	Ar	g	Asp 330	Asn	. Ph	e I	le	Gly	7 L 3	eu 35	Met	t
Tyr	Tyr	Asr	Ser 340	Th:	r Gl	уА	sn	Met	G:	l y 45	Val	Lev	ı Al	a (Sly	Gl: 35	n A	la	Se	r
Gln	Leu	Asr 35	n Ala	a Va	l Va	1 A	sp	Leu 360	G.	ln	Asp	Arg	g As	n :	Thr 365	Gl	u I	eu	Se	r
Tyr	Glr. 370		ı Le	ı Le	u As	p S	Ser 375	Ile	e G	ly	Asp	Ar	g Th 38	r 30	Arg	Ту	r F	Phe	Se	r
Met 385		As:	n Gl	n Al	a Va 39	al <i>F</i> 90	qaA	Ser	T :	yr	Asp	9 Pr	o As 5	gp	Val	. Ar	g I	Ile	11 40	.e 10
Glu	a Ası	n Hi	s Gl	y Th 40	r G:	lu A	qeA	Glu	ı L	eu	Pro 410	As	n T	yr	Суѕ	s Ph	e :	Pro 415	Le	eu
Gl	y Gl	y Va	l Il 42	e As	sn Ti	hr (Glu	Th	r I	eu 125	Th	r Ly	s V	al	Lys	s Pr 43	0 30	Lys	Tł	ır
Gl	y Gl	n Gl 43	u As	n G	Ly T	rp (Glu	Ly:	s F O	\sp	Al	a Th	ır G	lu	Phe 44!	e Se 5	er	Asp	L	ys
As	n Gl 45		e Ar	g V	al G	ly	Asn 455	As	n l	Phe	Al	a Me	et G 4	lu 60	Il	e As	sn	Leu	A	sn
Al 46		n Le	eu Ti	rp A	rg A	sn 70	Phe	e Le	u i	Гуг	: Se	r As	sn I 75	le	Al	a L	eu	Tyr	L 4	eu 80
Pr	o As	p L	ys Le	eu L 4	ys T 85	'yr	Sei	r Pr	0	Sei	c As	n Va	al I	ys	Il	e S	er	Asp 495	A S	sn
Pr	o As	sn T	hr T; 5	yr A 00	.sp 1	yr	Me	t As	sn	Ly: 50	s Ai 5	g V	al V	/al	Al	.a P 5	ro 10	Gly	, I	eu
Va	al As		ys T 15	yr I	le i	Asn	Le	u GI 52	Гу 20	Al	a Ai	rg T	rp :	Ser	Le 52	eu <i>P</i> 25	sp	Ту	r N	1et
As			al A	sn I	?ro	Phe	As 53	n H: 5	is	Hi	s A	rg A	sn .	Ala 540	G]	ly I	Leu	Ar	g :	fyr
			let I	eu l	Leu	Gly 550	As	n G	ly	Ar	g T	yr V	/al 555	Pro) Pl	he I	His	Il	e (Gln 560

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Val	Pro	Gln	Lys	Phe 565	Phe	Ala	Ile	Lys	Asn 570	Leu	Leu	Leu	Leu	Pro 575	Gly
Ser	Tyr	Thr	Tyr 580	Glu	Trp	Asn	Phe	Arg 585	Lys	Asp	Val	Asn	Met 590	Val	Leu
Gln	Ser	Ser 595	Leu	Gly	Asn	Asp	Leu 600	Arg	Val	Asp	Gly	Ala 605	Ser	Ile	Lys
Phe	Asp 610	Ser	Ile	Cys	Leu	Tyr 615	Ala	Thr	Phe	Phe	Pro 620	Met	Ala	His	Asn
Thr 625	Ala	Ser	Thir	Leu	Glu 630	Ala	Met	Leu	Arg	Asn 635	Asp	Thr	Asn	Asp	Gln 640
Ser	Phe	Asn	Asp	Tyr. 645	Leu	Ser	Ala	Ala	Asn 650	Met	Leu	Tyr	Pro	Ile. 655	Pro
Ala	Asn	Ala	Thr 660	Asn	Val	Pro	Ile	Ser 665	Ile	Pro	Ser	Arg	Asn 670	Trp	Ala
Ala	Phe	Arg 675	Gly	Trp	Ala	Phe	Thr 680	Arg	Leu	Lys	Thr	Lys 685	Glü	Thr	Pro
Ser	Leu 690	Gly	Ser	Gly	Tyr	Asp 695	Pro	Tyr	Tyr	Thr	Tyr 700	Ser	Gly	Ser	Île
Pro 705	Tyr	Leu	Asp	Gly	Thr 710	Phe	Tyr	Leu	Asn	His 715	Thr	Phe	Lys	Lys	Val 720
Ala	Ile	Thr	Phe	Asp 725	Ser	Ser	Val	Ser	Trp 730	Pro	Gly	Asņ	Asp	Arg 735	Leu
Leu	Thr	Pro	Asn 740	Glu	Phe	Glu	Ile	Lys 745	Arg	Ser	Val	Asp	Gly 750	Glu	Gly
Tyr	Asn	Val 755	Ala	Gln	Cys	Asn	Met 760	Thr	Lys	Asp	Trp	Phe 765	Leu	Val	Gln
Met	Leu 770	Ala	Asn	Tyr	Asn	11e 775	Gly	Tyr	Gln	Gly	Phe 780	Tyr	Ile	Pro	Glu
Ser 785	Tyr	Lys	Asp	Arg	Met 790	Tyr	Ser	Phe	Phe	Arg 795	Asn	Phe	Gln	Pro	Met 800
Ser	Arg	Gln	Val	Val 805	Asp	Asp	Thr	Lys	Tyr 810	Lys	Asp	Ţyr	Gln	Gln 815	Val.
Gly	Ile	Leu	His 820		His	Asn	Asn	Ser 825	_	Phe	Val	Gly	Tyr 830	Leu	Ala
Pro	Thr	Met 835	Arg	Glu	Gly	Gln	Ala 840	Tyr	Pro	Ala	Asn	Phe 845	Pro	Tyr	Pro
Leu	11e 850	Gly	Lys	Thr	Ala	Val 855	Asp	Ser	Ile	Thr	Gln 860	Lys	Lys	Phe	Leu
Cys 865	Asp	Arg	Thr	Leu	Trp 870	Arg	Ile	Pro	Phe	Ser 875	Ser	Asn	Phe	Met.	Ser 880
Met	Gly	Ala	Leu	Thr 885	•	Leu	Gly	Gln	Asn 890		Leu	Tyr	Ala	Asn 895	Ser

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Ala	His	Ala	Leu 900	Asp	Met	Thr	Phe	Glu 905	Val	Asp	Pro	Met	Asp 910	Glu	Pro		
		915			Leu		920					320					
Arg	Pro 930	His	Arg	Gly	Val	Ile 935	Glu	Thr	Val	Tyr	Leu 940	Arg	Thr	Pro	Phe		
Ser 945	Ala	Gly	Asn	Ala	Gln 950	His											
(2)	INF	'AMAC	TION	FOR	SEQ	ID	NO:5	5:							•		
	(i	(A) L B) T C) S	ENGT YPE: TRAN	HARA H: 9 nuc DEDN	8 ba leic ESS:	se p aci dou	airs .d	3								
	(ii) MO	LECU	JLE I	YPE:	oth	er r	iucle	eic a	acid							
	(xi) SE	QUEN	ICE I	ESCF	RIPTI	ON:	SEQ	ID 1	10:55	5:						
GAA Glu 1	CTC Lev	GG <i>F</i>	A GGT	y Gly	A GGT Y Gly	GGA Gly	A ACT	r AG' r Se:	r TT: r Phe	5 01,	A CGO	g Gl	A GAG y Ası	C ATT P Ile 15	CGC Arg		4.8
RAA Asr		AAGT?	ACTG	GAT'	CAT	GAC '	CTA	GACT	TA À	TTAA	GGAT(C CA	AATA	A			91
(2)	IN!	FORM	OITA	N FO	R SE	Q ID	NO:	56:									
	(i) S	(A) (B)	LEN TYPE	CHAR GTH: : am LOGY	17 ino	amin acid	o ac l	ids								
	(i	i) M	OLEC	ULE	TYPE	: pe	ptic	ie									
	(x	i) S	EQUE	NCE	DESC	RIPT	ION:	SEÇ	D ID	NO:5	66:						
Gl	u Le	u Gl	.y G]	Ly Gl	y Gl 5	y Gl	y Th	nr Se	er Ph	ne G1 10	Ly Ai	cg G	ly As	sp Il	le Arg	ſ	

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WHAT IS CLAIMED IS:

- 1. A chimeric adenovirus coat protein comprising a nonnative amino acid sequence, wherein said chimeric adenovirus coat protein has a decreased ability or inability to be recognized by a neutralizing antibody directed against the wild-type adenovirus coat protein.
- 2. The chimeric adenovirus coat protein of claim 1, wherein said nonnative amino acid sequence comprises a deletion, insertion, or a replacement of a region of from about 1 to about 750 amino acids of said wild-type adenovirus coat protein.
- 3. The chimeric adenovirus coat protein of claim 1 or 2, wherein said nonnative amino acid sequence comprises a plurality of deletions, insertions, and/or replacements.
- 4. The chimeric adenovirus coat protein of any of claims 1-3, wherein said coat protein is a chimeric adenovirus hexon protein.
- 5. The chimeric adenovirus coat protein of claim 4, wherein said region deleted or replaced comprises a hypervariable region in either the 11 loop or the 12 loop.
- 6. The chimeric adenovirus coat protein of claim 5, wherein said hypervariable region is selected from the group consisting of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and HVR7.
- 7. The chimeric adenovirus coat protein of any of claims 1-6, comprising a sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID

- NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.
- 8. The chimeric adenovirus coat protein of any of claims 1-7, wherein said nonnative amino acid sequence comprises a spacer of about 1 to about 750 amino acids.
- 9. The chimeric coat adenovirus coat protein of claim 8, wherein said spacer comprises the sequence of SEQ ID NO:50.
- 10. The chimeric adenovirus coat protein of any of claims 1-9, comprising an amino acid sequence of a coat protein of another serotype of adenovirus.
- 11. The chimeric adenovirus coat protein of claim 10, wherein said coat protein of another serotype is a hexon protein.
- 12. An isolated or purified nucleic acid that encodes the chimeric adenovirus coat protein of any of claims 1-11.
- 13. The isolated or purified nucleic acid of claim
 12 comprising a sequence selected from the group
 consisting of SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ
 ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID
 NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID
 NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID
 NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID
 NO:43, SEQ ID NO:45, and SEQ ID NO:47.

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- 14. The isolated or purified nucleic acid of claim 12 or 13 comprising SEQ ID NO:49.
- 15. An adenoviral vector that comprises the chimeric adenovirus coat protein of any of claims 1-11.
- 16. A method of genetically modifying a cell which comprises contacting said cell with the adenoviral vector of claim 15.
- 17. A host cell that comprises the chimeric adenovirus coat protein of any of claims 1-11.
- 18. A method of constructing an adenoviral vector that has a decreased ability or inability to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein, which method comprises obtaining an adenoviral vector comprising a wild-type adenovirus coat protein and replacing said wild-type adenovirus coat protein with the chimeric adenovirus coat protein of any of claims 1-11.

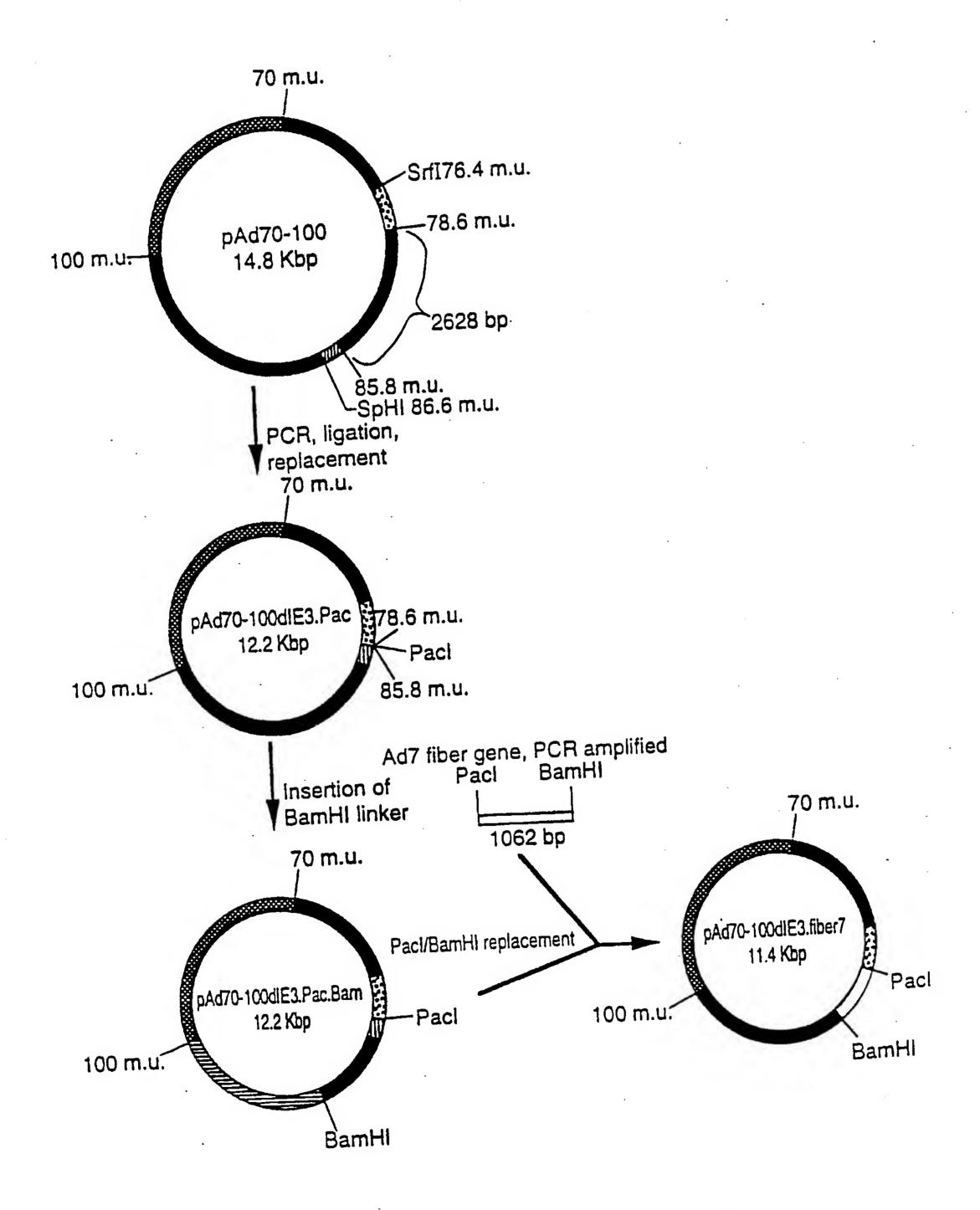
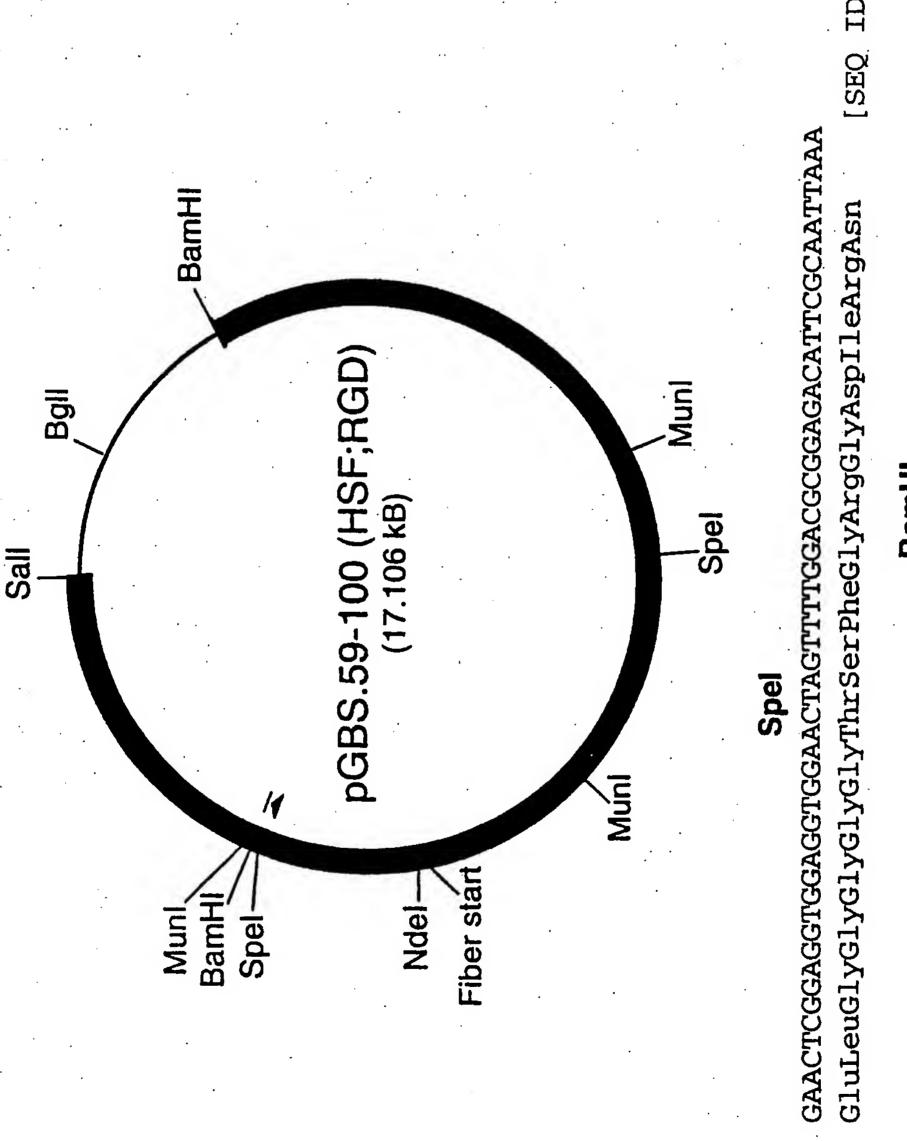


FIG. 1
SUBSTITUTE SHEET (RULE 26)



ID NO:56] ID NO:55] [SEQ GluLeuGlyGlyGlyGlyThrSerPheGlyArgGlyAspIleArgAsn GTACTGGATTCATGACTCTAGACTTAATTTAAGGATCCAATAAA BamHI

FIG.

INTERNATIONAL SEARCH REPORT

Int .tional Application No PCT/US 98/05033

A. CLASSIFIC	CATION OF SUBJECT MATTER C12N15/86 C07K14/075 C12N15/34	C12N5/10	
According to It	nternational Patent Classification (IPC) or to both national classificati	ion and IPC	
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Minimum docu	umentation searched (classification system followed by classification C12N C07K	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	
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Documentatio	on searched other trials trialmost described		
Electronic dal	ta base consulted during the international search (name of data bas	e and, where practical, search terms used)	
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C. DOCUME	NTS CONSIDERED TO BE RELEVANT		Dalaman to alaim No
Category °	Citation of document, with indication, where appropriate, of the rele	evant passages	Relevant to claim No.
A	WO 96 26281 A (GENVEC, INC.) 29 /	August	1-18
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	see table 1		
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X Fu	rther documents are listed in the continuation of box C.	X Patent family members are liste	d in annex.
• Special	categories of cited documents :	"T" later document published after the in	nternational filing date
"A" docum	ment defining the general state of the art which is not sidered to be of particular relevance	cited to understand the principle of invention	theory underlying the
"E" earlie filling	r document but published on or after the international g date	"X" document of particular relevance; the cannot be considered novel or car involve an inventive step when the	document is taken alone
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	r than the priority date claimed ne actual completion of theinternational search	Date of mailing of the international	
Date of the	9 July 1998	27/07/1998	
		Authorized officer	
Name ar	nd mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Cupido, M	

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